

# Role of Radiation Therapy in TIMEs: Good, Bad, Don't Know

**Puneeth Iyengar, MD, PhD**  
**Attending**  
**Director, Metastatic Service**  
**Member, Thoracic Service**  
**Department of Radiation Oncology**  
**Memorial Sloan Kettering Cancer Center**

**Member, Weill Center for Metabolism**

**Adjunct Faculty, UT Southwestern Medical Center**

**MATOS Masters of Thoracic Oncology 2023**  
**Albuquerque, New Mexico**

4K ULTRAHD™

CLINT EASTWOOD



**THE GOOD THE BAD and THE UGLY**

co-starring  
**LEE VAN CLEEF**

also starring  
**ELI WALLACH**  
in the role of TUCO

directed by  
**SERGIO LEONE**

Would make the argument that each of these characters had a little bit of good, bad, and ugly in them...**dependent on the context and their position.**

Same applies for radiation and its effect on TIMEs.



Memorial Sloan Kettering  
Cancer Center

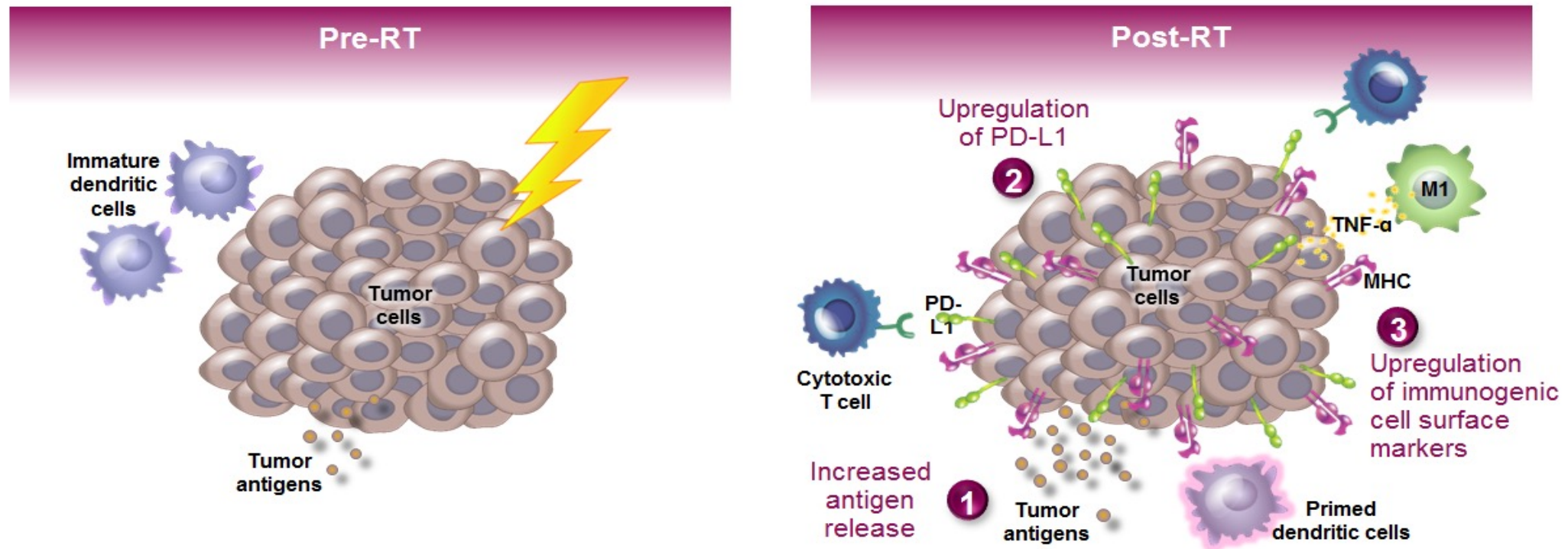
# How to integrate radiotherapy in the modern and rapidly-changing era?

- Emphasis on multimodality therapy
- **Sequence and timing of radiation therapy may be critically important**
- Variation may depend on heterogeneity in NSCLC
- Molecular considerations may impact response
- Importance of clinical trials to tease all of this out



# Immunotherapy biomarkers are upregulated following RT

RT Induces Multiple Immunomodulatory Changes That May Influence the Effectiveness of Immunotherapy<sup>1-3</sup>



M1, tumor-associated macrophage; MHC I, major histocompatibility complex I; PD-L1, programmed cell death-ligand 1; TNF- $\alpha$ , tumor necrosis factor alpha.

1. Daly ME, et al. *J Thorac Oncol.* 2015;10(12):1685-1693. 2. Kaur P, Asea A. *Frontiers Oncol.* 2012;2:191. 3. Deng L, et al. *J Clin Invest.* 2014;124(2):687-695.



[nature](#) > [nature reviews cancer](#) > [review articles](#) > article

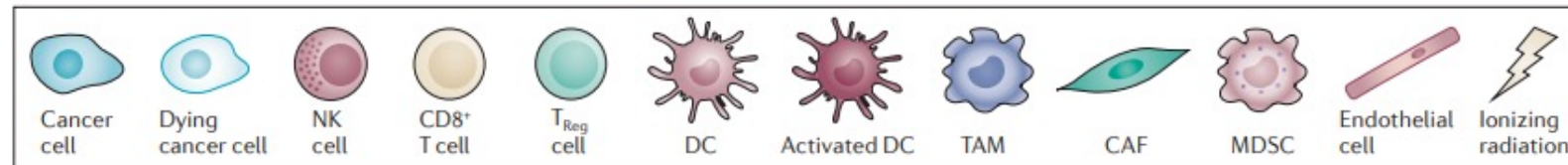
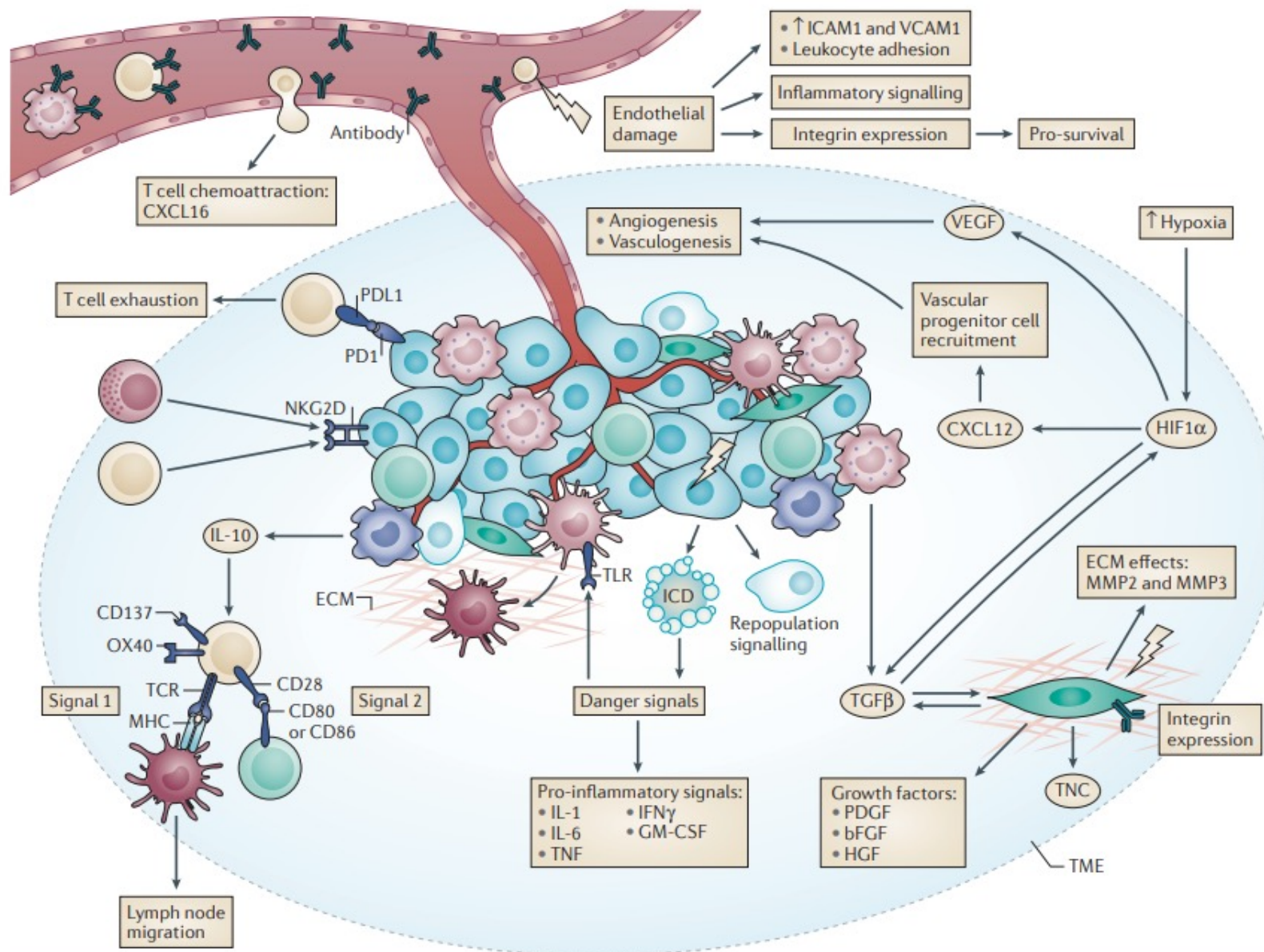
[Published: 24 June 2015](#)

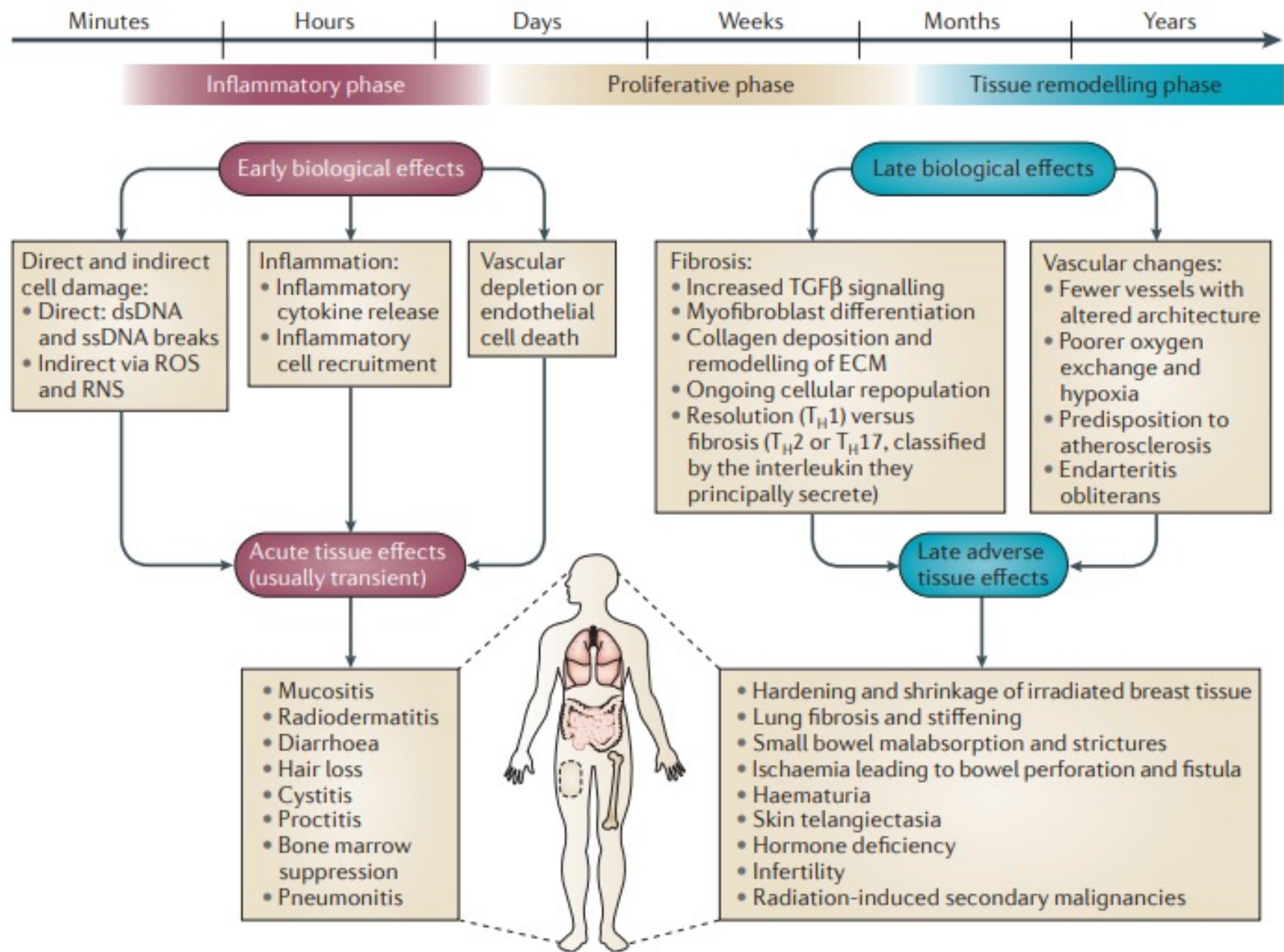
# The tumour microenvironment after radiotherapy: mechanisms of resistance and recurrence

[Holly E. Barker](#) , [James T. E. Paget](#), [Aadil A. Khan](#) & [Kevin J. Harrington](#)

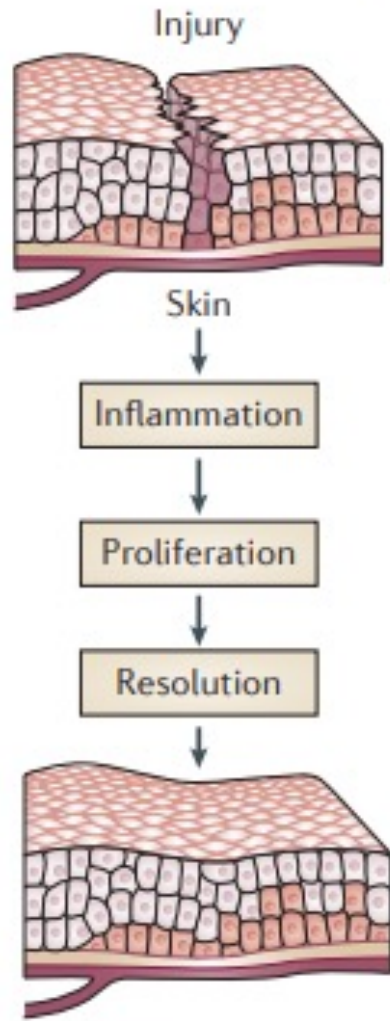
[Nature Reviews Cancer](#) **15**, 409–425 (2015) | [Cite this article](#)

**33k** Accesses | **1248** Citations | **45** Altmetric | [Metrics](#)

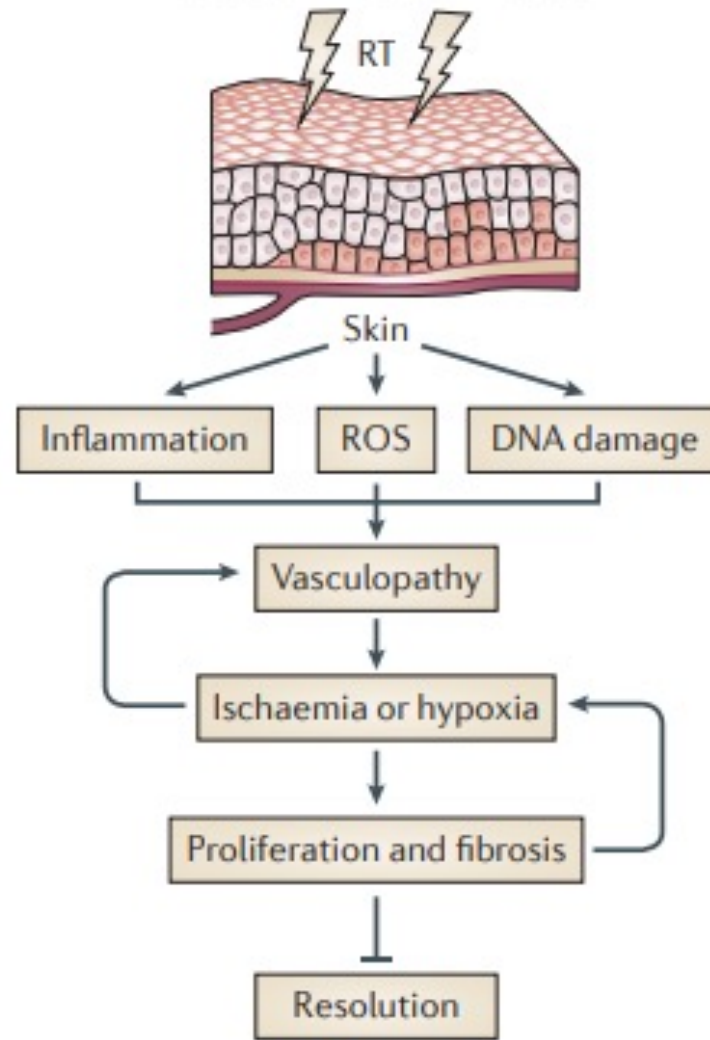




### Normal wound healing



### Radiation-induced fibrosis





Please add cGAS/STING pathways to this mix.

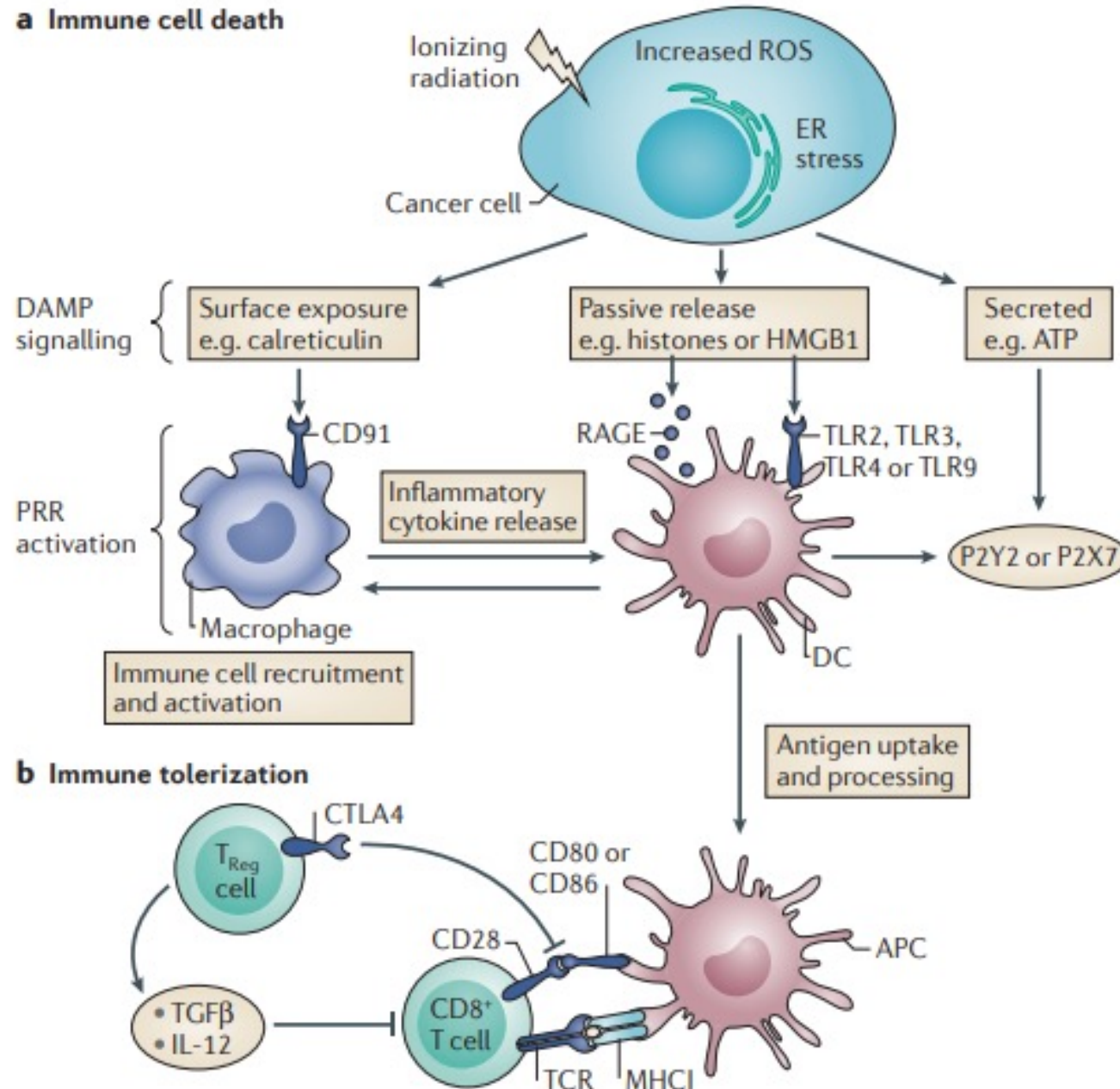


Table 1 | Current and future TME targets for radiosensitization

Resistance mechanism	Drugs	Targets	Mode of action	
Immune response	Ipilimumab	CTLA4	T cell activation	
	Nivolumab and pembrolizumab	PD1	T cell exhaustion	
	Imiquimod	TLR7	DC activation	
	Oncolytic viruses	Tumour cells	Activate immune response	
	Future inhibitors	IL-6 and IL-10	PDL1, TIM3 and LAG3	Prevent T cell exhaustion
			GM-CSF, CXCL16, OX40, CD40L, CD80 and CD137	T cell recruitment and activation
	Future agonists	CCL3, CCL5, IL-2, IL-4, IL-12 and IRX-2	Activate immune response	
Hypoxia	Nitroimidazole derivatives (that is, nimorazole)	Hypoxic cells	Reduce tumour hypoxia	
	Bioreactive albumin-MnO <sub>2</sub> nanoparticles	Hypoxic cells	Reduce tumour hypoxia	
	Acriflavine and YC-1	HIF1 $\alpha$	Reduce hypoxia response pathway activity	
	Aflibercept	All VEGF molecules and PlGF	Vessel normalization	
	AMG386	ANG1 and ANG2	Inhibit pBMDc recruitment	
	Endostar	VEGF, TGF $\beta$ , HIF1 $\alpha$ and bFGF	Inhibit angiogenesis	
	AMD3100	CXCL12 and CXCR4	Inhibit BMDc recruitment and vasculogenesis	
	Integrin inhibitors (cilengitide, vitaxin and volociximab)	Integrins $\alpha$ v $\beta$ 3, $\alpha$ v $\beta$ 5 and $\alpha$ 5 $\beta$ 1	Inhibit angiogenesis	
	Future inhibitors	Integrins $\alpha$ 6 $\beta$ 1 and $\alpha$ 6 $\beta$ 4	Reduce endothelial cell survival and inhibit angiogenesis	
	Future inhibitors	PlGF and ANG2	Vessel normalization and overcome resistance to anti-VEGF therapies	
Fibrotic processes	BIBF1000 and BIBF1120	PDGF, VEGF and bFGF receptors	Reduce GF signalling and TME remodelling; fibrosis	
	Imatinib, nilotinib and dasatinib	TGF $\beta$ and PDGF	GF signalling; collagen synthesis	
	Vismodegib, saridegib and sonidegib	SMO	Reduce HH signalling; fibrosis	
	Suramin	PDGF, EGF, TGF $\beta$ , FGF2 and IGF receptors and heparanase enzymes	Reduce GF signalling and TME remodelling; fibrosis	
	ST0001, PG545, M402 and PI-88	Heparanase	Inhibit TME remodelling	
	SD-208	TGF $\beta$ R1	Inhibit TGF $\beta$ signalling	
	Simtuzumab	LOXL2	Reduce TME remodelling; liver fibrosis	
	81C6 and F16SIP	TNC	Reduce CAF-mediated TME remodelling	
	Future inhibitors	HGF, CTGF, MMP2, MMP3, and integrins $\alpha$ 11 $\beta$ 1, $\alpha$ v $\beta$ 6 and $\alpha$ 3 $\beta$ 1	TME activation and remodelling; radiation-mediated fibrosis	

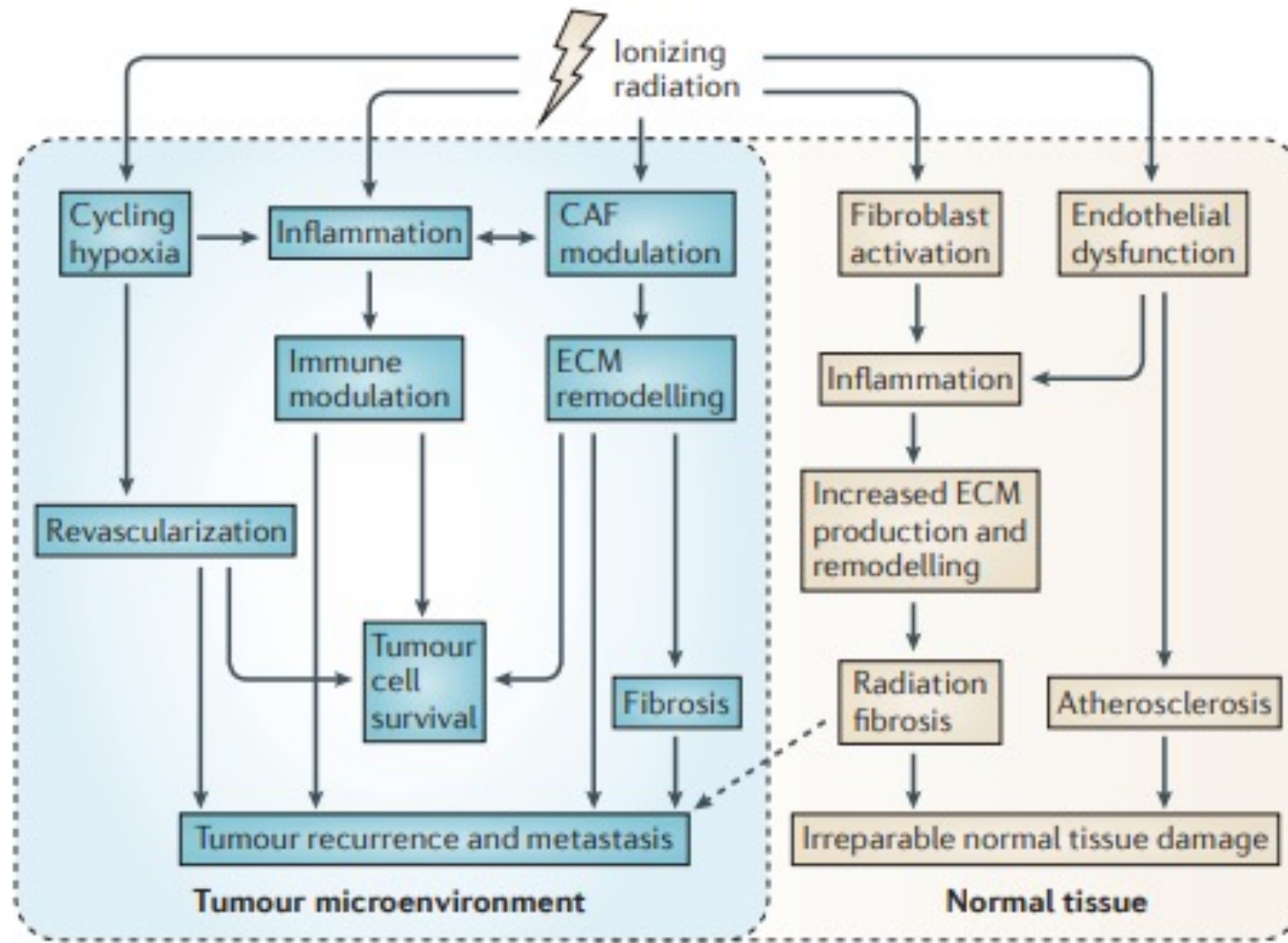


Table 2 | **Fractionation of radiotherapy and timing of immunomodulation**

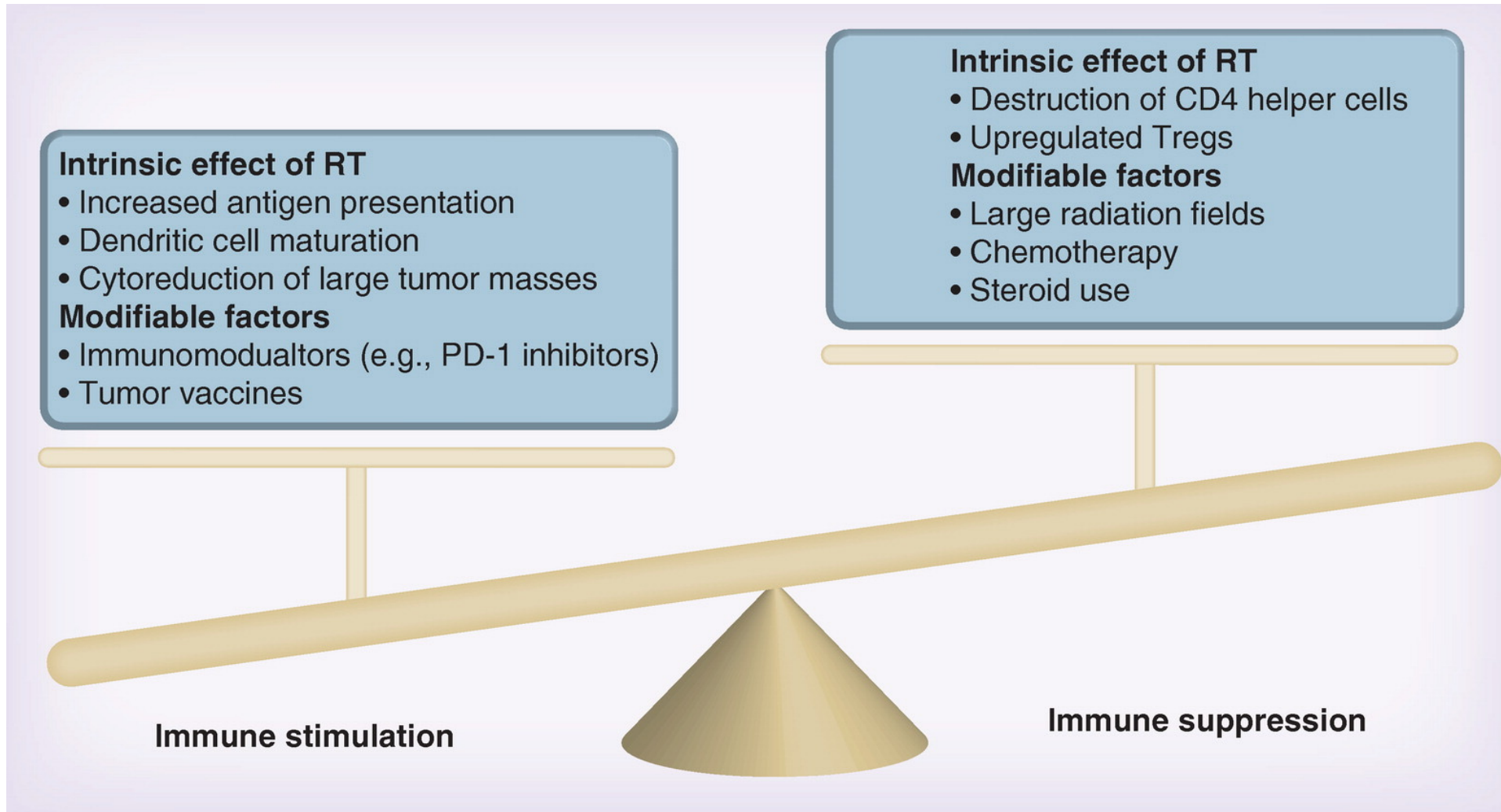
Study type	Cancer type	Intervention	Outcome	Refs
<i>Fractionation</i>				
Preclinical	Murine glioma	<ul style="list-style-type: none"> <li>• RT: 10 Gy in one fraction</li> <li>• Immunomodulation: anti-PD1</li> </ul>	Longer survival seen (53 days (RT and anti-PD1) versus 25 days (control), 27 days (anti-PD1) or 28 days (RT))	190
Preclinical	Murine breast cancer	<ul style="list-style-type: none"> <li>• RT: 20 Gy in one fraction, 24 Gy in three fractions or 30 Gy in five fractions</li> <li>• Immunomodulation: anti-CTLA4</li> </ul>	Best response seen in 24 Gy in three fractions	171
Preclinical	Murine breast cancer	<ul style="list-style-type: none"> <li>• RT: 12 Gy in one fraction, 24 Gy in two fractions</li> <li>• Immunomodulation: anti-CTLA4</li> </ul>	CD8 <sup>+</sup> T cell antitumour immunity demonstrated	191
Clinical	Patients with hepatoma	<ul style="list-style-type: none"> <li>• RT: 8 Gy in one fraction</li> <li>• Immunomodulation: DC vaccine</li> </ul>	Two partial responses, four minor ones	192
Clinical	Prostate cancer	<ul style="list-style-type: none"> <li>• RT: 70 Gy in 30 fractions</li> <li>• Immunomodulation: IL-2 and GM-CSF</li> </ul>	Increased levels of PSA-specific T cells	193
Clinical	MF	<ul style="list-style-type: none"> <li>• RT: 9–18 Gy in nine fractions</li> <li>• Immunomodulation: injected TLR9 agonist</li> </ul>	5 out of 15 responses	194
<i>Timing</i>				
Clinical	MM or RCC	<ul style="list-style-type: none"> <li>• RT: 60 Gy in three fractions</li> <li>• Immunomodulation: IL-2</li> </ul>	8 out of 12 responses	195
Preclinical	Murine prostate cancer expressing HA	<ul style="list-style-type: none"> <li>• RT: 15 Gy in one fraction</li> <li>• Immunomodulation: CD4<sup>+</sup> T cells primed against HA (given at points on a time course after RT)</li> </ul>	Tolerance seen between days 3 and 16. Normal T cell response by day 33	55
Clinical case report	MM	<ul style="list-style-type: none"> <li>• RT: 28.5 Gy in three fractions</li> <li>• Immunomodulation: ipilimumab (anti-CTLA4) at 1 month after RT</li> </ul>	Complete response	74
Clinical case report	MM	<ul style="list-style-type: none"> <li>• RT: 54 Gy in three fractions</li> <li>• Immunomodulation: ipilimumab</li> </ul>	Complete response	54

CTLA4, cytotoxic T lymphocyte antigen 4; DC, dendritic cell; HA, haemagglutinin antigen; MF, mycosis fungoides; MM, malignant melanoma; PD1, programmed cell death protein 1; PSA, prostate-specific antigen; RCC, renal cell carcinoma; RT, radiotherapy; TLR, Toll-like receptor.





# Balance immune-stimulatory effects and suppressive effects of RT



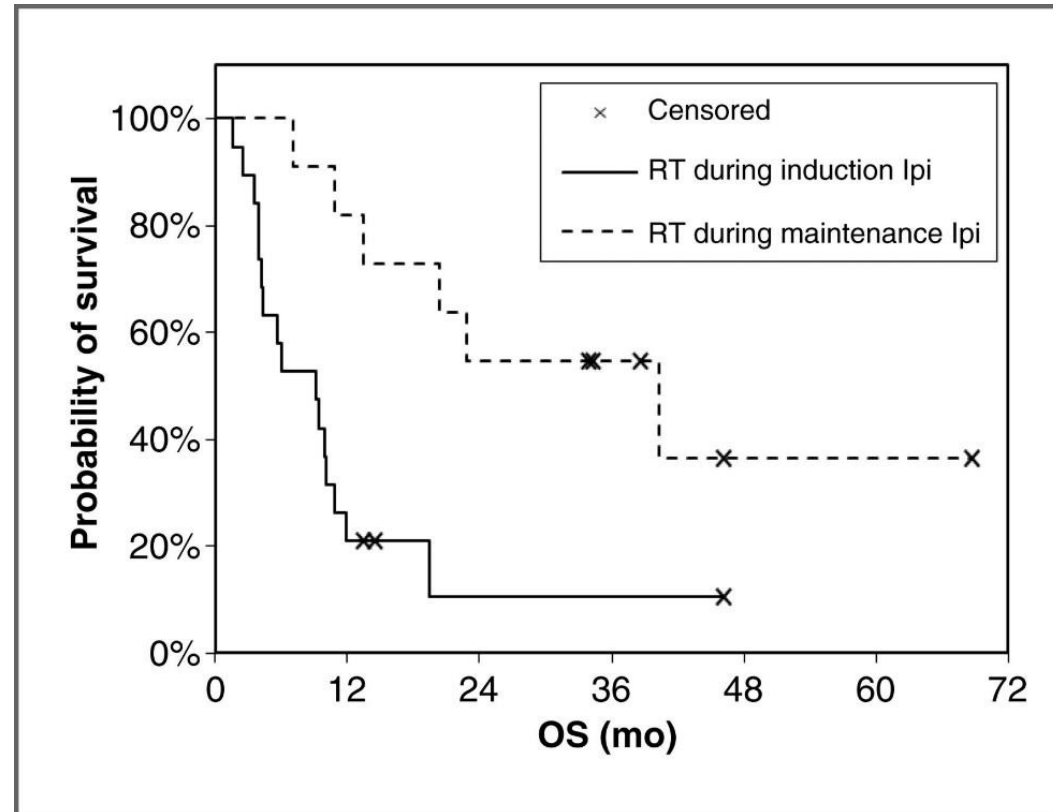
# Potential Benefits of Combining RT and Immunotherapy

- SBRT is less immunosuppressive than conventionally fractionated RT or sx
  - SBRT specifically can even be immunostimulatory and deplete immunosuppressive cells
- RT can improve antigen presentation by antigen presenting cells
  - SBRT specifically can release high levels of tumor antigens
- SBRT upregulates immunogenic cell surface markers (ie. MHC-1)
- SBRT can induce immunogenic cell death
- RT and especially SBRT can increase homing of immune cells to tumor
- RT can recruit regulatory T cells (Tregs)
- RT can shift tumor-associated macrophages polarization from M2 to M1
- RT can induce secretion of danger signals and cytokines (ie. TNFalpha)
- RT can upregulate cell-surface expression of PD-L1



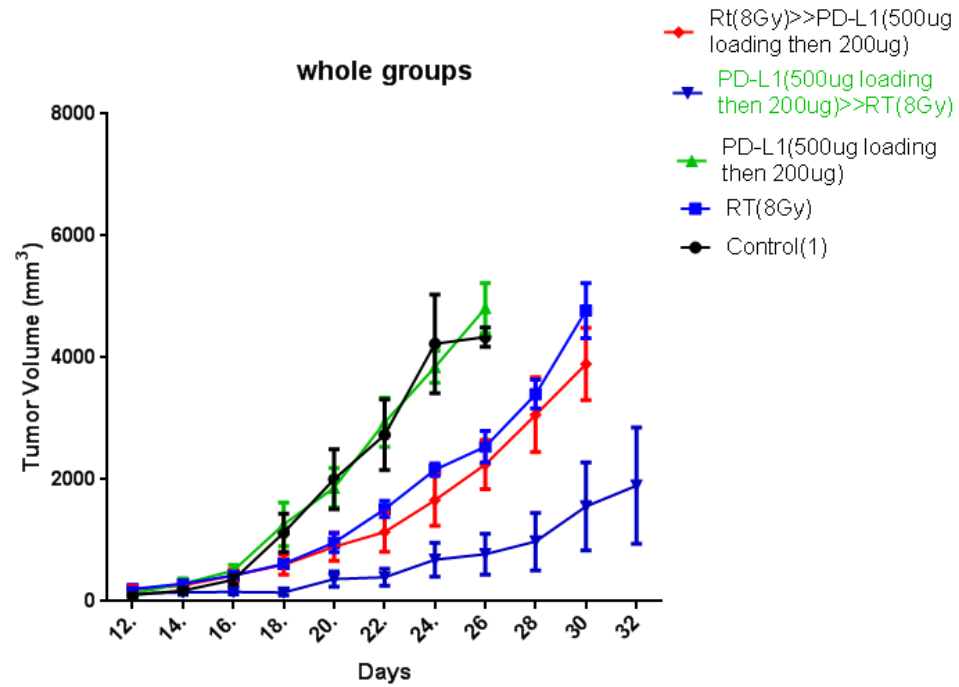
# RT + Immunotherapy: The Importance of Timing

- MSKCC retrospective study of melanoma patients treated with ipilimumab and extracranial RT
- Median OS: 9 months when RT given during induction vs. 39 months when RT given during maintenance



Barker CA, et al. *Cancer Immunol Res.* 2013;1(2):92-98.

# Timing of Immunotherapy and SBRT



Significantly superior tumor control was achieved in Balb/c mice when the PD-L1 blockade was delivered prior to radiotherapy to 8 Gy



# PACIFIC: Study Design

Phase 3, Randomized, Double-blind, Placebo-controlled, Multicenter, International Study

- Unresectable, Stage III NSCLC without progression after definitive platinum-based cCRT ( $\geq 2$  cycles)
- 18 years or older
- WHO PS score 0 or 1
- If available, archived pre-cCRT tumor tissue for PD-L1 testing\*

**All-comers population  
(i.e. irrespective of PD-L1 status)**

**N=713 randomized**

1–42 days  
post-cCRT

**R**

**Durvalumab**  
10 mg/kg q2w for  
up to 12 months  
N=476

2:1 randomization,  
stratified by age, sex, and  
smoking history

**Placebo**  
10 mg/kg q2w for  
up to 12 months  
N=237

## Primary endpoints

- PFS by BICR using RECIST v1.1†
- OS

## Key secondary endpoints

- ORR, DoR and TTDM by BICR
- PFS2 by investigator
- Safety
- PROs



# PACIFIC: Prognostic baseline factors for OS (ITT)

Baseline Variable	Comparator		Reference		HR (95% CI)
	Group	No. of Events/Total No. of Patients (%)	Group	No. of Events/Total No. of Patients (%)	
Treatment arm	Durvalumab	264/476 (55.5)	Placebo	155/237 (65.4)	0.71 (0.58 to 0.87) <sup>a</sup>
Age, years	≥ 65	210/322 (65.2)	< 65	209/391 (53.5)	1.30 (1.06 to 1.59) <sup>a</sup>
Disease stage <sup>b</sup>	IIIB	182/319 (57.1)	IIIA	227/377 (60.2)	1.03 (0.84 to 1.26)
Best response to prior treatment <sup>c</sup>	CR/PR	195/365 (53.4)	SD	216/338 (63.9)	0.88 (0.72 to 1.08)
Tumor histologic type	Squamous	205/326 (62.9)	Nonsquamous	214/387 (55.3)	1.28 (1.04 to 1.58) <sup>a</sup>
WHO PS	1 <sup>d</sup>	233/365 (63.8)	0	186/348 (53.4)	1.23 (1.01 to 1.50) <sup>a</sup>
Prior platinum CT agent <sup>e</sup>	Cisplatin	215/395 (54.4)	Carboplatin	190/301 (63.1)	0.84 (0.69 to 1.03)
Race	Asian	95/192 (49.5)	White	310/494 (62.8)	0.63 (0.49 to 0.81) <sup>a</sup>
	Black or African American	7/14 (50.0)			0.81 (0.38 to 1.73)
	Other <sup>f</sup>	7/13 (53.8)			0.91 (0.41 to 1.99)
Sex	Male	304/500 (60.8)	Female	115/213 (54.0)	1.27 (1.01 to 1.61) <sup>a</sup>
Smoking status	Smoker	384/649 (59.2)	Nonsmoker	35/64 (54.7)	0.83 (0.56 to 1.22)
Time from CRT to random assignment, days	≥ 14	312/531 (58.8)	< 14	107/182 (58.8)	0.97 (0.77 to 1.22)
EGFR or ALK aberration status	Positive <sup>g</sup>	25/43 (58.1)	Negative	275/482 (57.1)	1.06 (0.69 to 1.64)
	Unknown	119/188 (63.3)			0.95 (0.73 to 1.23)
PD-L1 expression level	TC ≥ 25%	78/159 (49.1)	TC < 25%	175/292 (59.9)	0.82 (0.62 to 1.07)
	Unknown	166/262 (63.4)			1.19 (0.92 to 1.54)



Spigel, D JCO, December 2021.

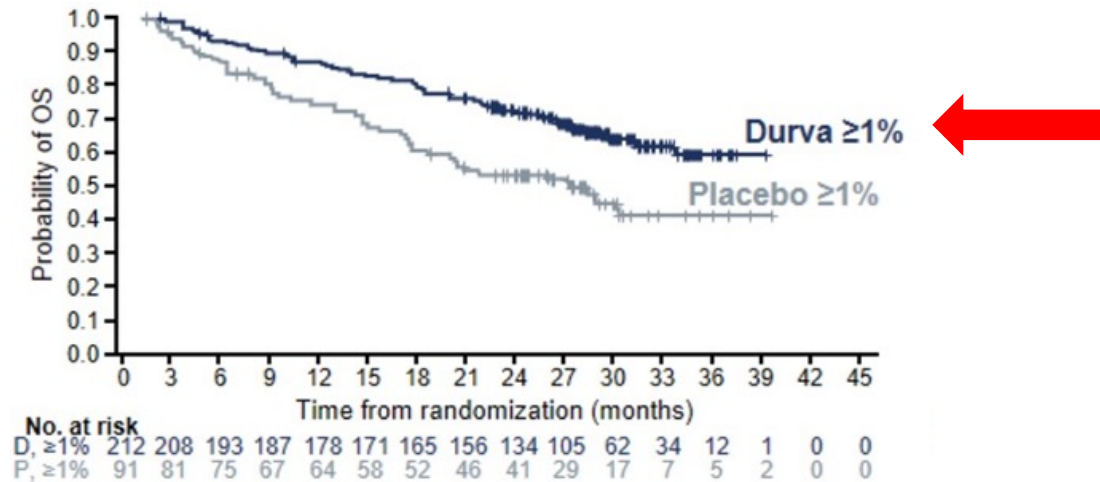


Memorial Sloan Kettering  
Cancer Center

# Conclusions on Outcomes by PD-L1 Status is not definitive due to limitations

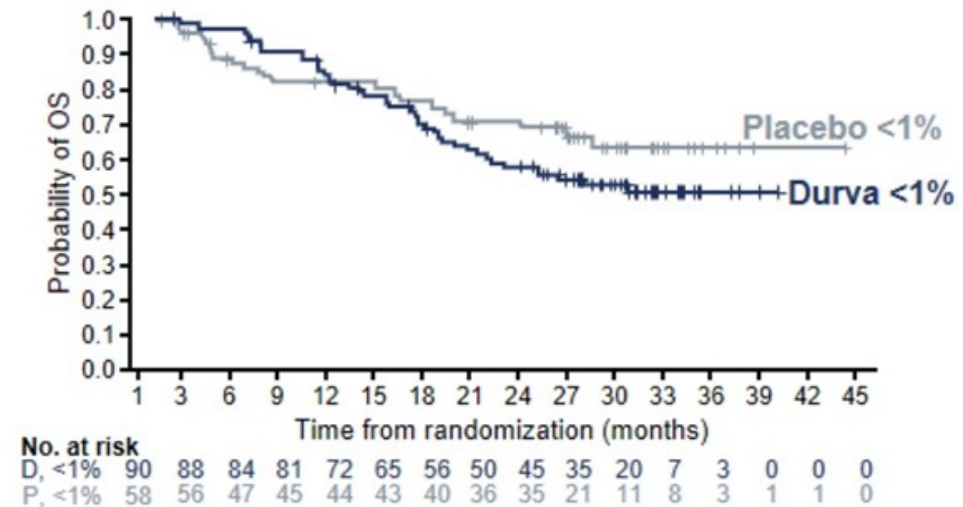
## OS (BICR) by PD-L1 TC $\geq 1\%$

	No. events / no. patients (%)	Median OS (95% CI), mo
Durvalumab, $\geq 1\%$	70/212 (33.0)	NR (NR, NR)
Placebo, $\geq 1\%$	45/91 (49.5)	29.1 (17.7, NR)
$\geq 1\%$ OS HR 0.53 (95% CI 0.36, 0.77)		



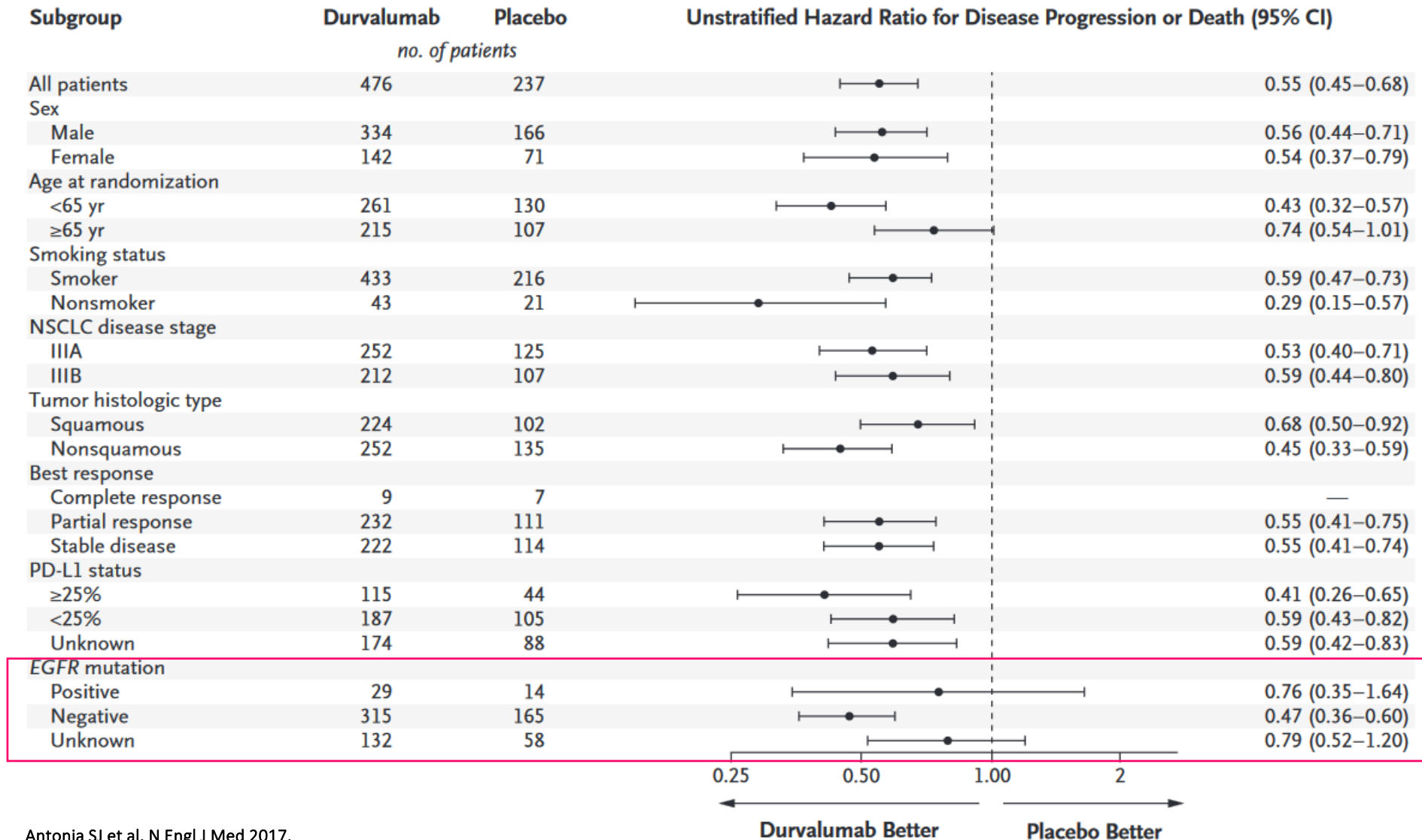
## OS (BICR) by PD-L1 TC $< 1\%$

	No. events / no. patients (%)	Median OS (95% CI), mo
Durvalumab, $< 1\%$	41/90 (45.6)	NR (20.8, NR)
Placebo, $< 1\%$	19/58 (32.8)	NR (27.3, NR)
$\geq 1\%$ OS HR 1.36 (95% CI 0.79, 2.34)		



- ◆ In the PD-L1 TC  $< 1\%$  subgroup, imbalances exist in baseline characteristics.
- ◆ Placebo arm: > more males, SQCLC, and Stage IIIB.

# DFS by EGFR status



# Consolidation Durvalumab for Stage III EGFRmut NSCLC — Stanford, City of Hope, UCSF, UC Davis

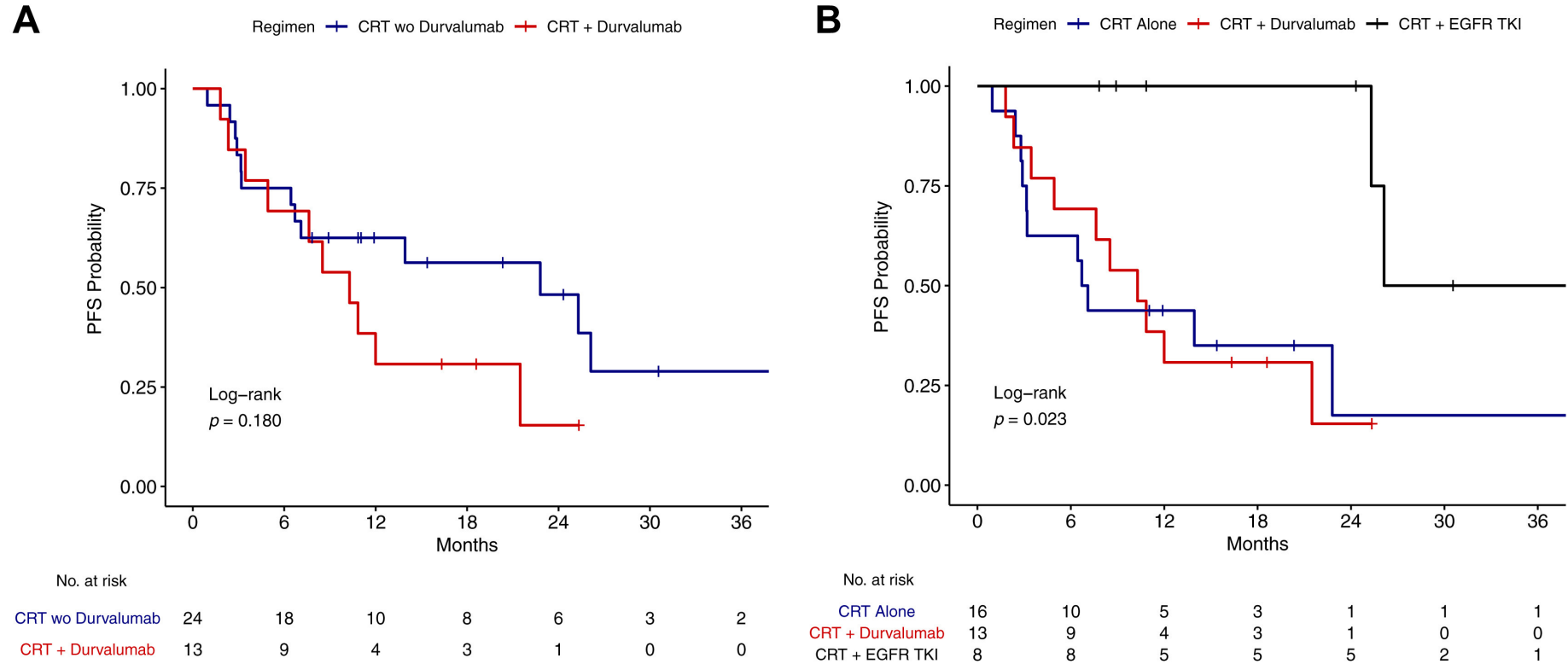


Figure 3: PFS after chemoXRT +/- Durva

(A) Median PFS CRT + durvalumab versus CRT wo durvalumab  
10.3 months versus 22.8 months (log-rank  $p = 0.180$ ).

(B) Median PFS CRT alone versus CRT durvalumab versus CRT + EGFR TKI :  
6.9 mo vs 10.3 mo vs 26.1 mon (log-rank  $p = 0.023$ ).

# Conclusions

**Radiation therapy effects on TIMEs and, as a consequence, tumor control can be influenced by:**

1. Sequencing of systemic therapy and local therapy
2. Type of systemic therapy and type of radiation
3. Dose of radiation
4. The understudied variation in host immune and systemic biology responses to tumor and therapy (systemic therapy and/or radiation)

