# Cancer Immunotherapy: Current and Next Generation Biomarkers

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# **Definition of Cancer Immunotherapy**

- Treatments that harness patients' immune system for cancer therapy
- Monoclonal antibodies
  - Target cancer cells
  - Modulate immune function: Immune Checkpoint Blockade (ICB)
- Cytokines (also blocking antibodies)
- Cancer vaccines (+/- dendritic cells)
- Adoptive cellular therapy (+/- engineered receptors: TCR or chimeric antigen receptor, CAR)

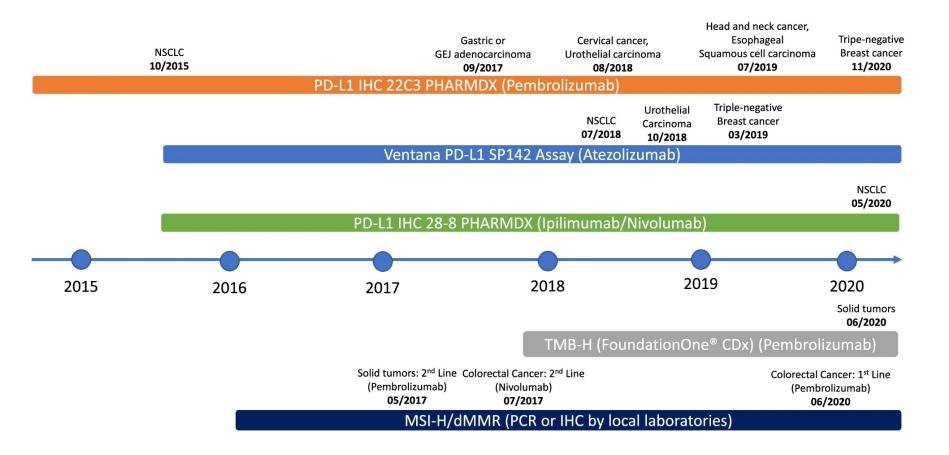
# Approved Immune Checkpoint Inhibitors (ICIs) for Cancer

- 2011: Ipilimumab (anti-CTLA4 Ab) for advanced melanoma
- 2014: Pembrolizumab and Nivolumab (anti-PD1 Ab) for advanced melanoma
- 2015: Nivolumab for lung cancer and kidney cancer; Nivo/Ipi combination for melanoma
- 2016 on: multiple other cancer types; MSI tumors

These treatments still only work for subsets of patients and some cancer types (20-30% of hot tumors)

#### **Need for biomarkers!**

#### Current FDA Approved Biomarkers for ICIs



Sankar et al. Biomarker Research 2022

#### Current FDA Approved Biomarkers for ICIs

Biomarker	Cancer types	ICI agents	Accuracy
Programmed death- ligand 1 (PD-L1)	NSCLC, gastric, cervical, urothelial, HNSCC, esophageal, TNBC	Pembrolizmab Ipilimumab/Nivolumab Atezolizmab	low diagnostic accuracy: predictive of only 28.9% of ICI approvals 2011- 2019.
Microsatellite instability/defective mismatch repair (MSI/dMMR)	Any (<5% of metastatic tumors): mainly CRC, endometrial, ovarian, gastric, other GI	Pembrolizmab Nivolumab	Good, but also depends on method: IHC, PCR, NGS
Tumor mutational burden (TMB)	Any: include CRC, esophageal, NSCLC, SCLC	Pembrolizmab	Wide variation in results, also depends on method: WES, targeted NGS panels

#### FDA Approved PD-L1 IHC Assays

Test Name	PMA#	Tumor Type	ICI	Approval Year	Scoring System	PD-L1-Threshold	PD-L1 Staining
PD-L1 IHC 22C3 pharmDx	P150013	NSCLC	Pembrolizumab	2015	TPS	>=50%	tumor cells
PD-L1 IHC 22C3 pharmDx	P150013/ S006	gastric or GEJ adenocarcinoma	Pembrolizumab	2017	CPS	>=1	tumor cells, lymphocytes, macrophages
PD-L1 IHC 22C3 pharmDx	P150013/ S009	Cervical Cancer	Pembrolizumab	2018	CPS	>=1	tumor cells, lymphocytes, macrophages
PD-L1 IHC 22C3 pharmDx	P150013/ S011	urothelial carcinoma	Pembrolizumab	2018	CPS	>=10	tumor cells, lymphocytes, macrophages
PD-L1 IHC 22C3 pharmDx	P150013/ S014	head and neck squamous cell carcinoma	Pembrolizumab	2019	CPS	>=1	tumor cells, lymphocytes, macrophages
PD-L1 IHC 22C3 pharmDx	P150013/ S016	esophageal squamous cell carcinoma	Pembrolizumab	2019	CPS	>=10	tumor cells, lymphocytes, macrophages
VENTANA PD-L1 (SP142) Assay	P160002/ S006	urothelial carcinoma/ NSCLC	atezolizumab	2018	IC%/IC% or TPS	>=5%/>=10% or >=50%	tumor area/tumor area, tumor ells
VENTANA PD-L1 (SP142) Assay	P160002/ S009	Triple-Negative Breast Carcinoma	atezolizumab	2019	IC%	>=1%	tumor area
VENTANA PD-L1 (SP142) Assay	P160002/ S012	NSCLC	atezolizumab	2020	IC%/TPS	>=10%/>=50%	tumor area Tumor cells
PD-L1 IHC 28-8 pharmDx	P150025/ S013	NSCLC/SCCHN/UC	Nivolumab in combination with ipilimumab	2020	TPS	>=1%	tumor cells
PD-L1 IHC SP263	P160046	urothelial carcinoma	Durvalumab	2017	TPS/ICP/ IC+	>=25%/ICP > 1% and IC+ >=25%/ICP = 1% and IC+ = 100%.	tumor cells Immune cells

Wang et al. Frontiers in Oncology 2021

#### Tumor PD-L1 is a borderline predictor

#### (A) pfs

	PD-L1 po	sitive	PD-L1 neg	ative		Odds Ratio	Odds	Ratio	
Study or Subgroup	Events	Total	Events	Total	Weight	M-H. Fixed, 95% C	M-H, Fix	ed. 95% CI	
Adams1 2018 (31)	9	105	5	64	15.8%	1.11 [0.35, 3.46]			
Adams3 2018 P	3	12	2	12	4.2%	1.67 [0.22, 12.35]		· · · · · · · · · · · · · · · · · · ·	
Emens 2018 <sup>[10]</sup>	10	91	1	21	4.0%	2.47 [0.30, 20.43]		· · · · · · · · · · · · · · ·	
Schmid 2018 <sup>(36)</sup>	38	185	39	266	70.8%	1.50 [0.92, 2.46]		-	
Sherene 2019 <sup>(29)</sup>	4	40	0	12	1.9%	3.08 [0.15, 61.38]		· · · • · · · · · · ·	
Voorwerk 2019 <sup>[12]</sup>	5	44	1	21	3.3%	2.56 [0.28, 23.46]			
Total (95% CI)		477		396	100.0%	1.55 [1.02, 2.36]		<b>٠</b>	
Total events	69		48						
Heterogeneity: Chi <sup>2</sup> = 0	).94, df = 5 (	P = 0.9	7); 12 = 0%				0.001 0.1	1 10	1000
Test for overall effect: 2	Z = 2.07 (P	= 0.04)					Favours (PD-L1 negative)		

#### (B) os

	PD-L1 po	sitive	PD-L1 neg	ative		Odds Ratio	Odds Ratio	
Study or Subgroup	Events	Total	Events	Total	Weight	M-H. Fixed, 95% C	M-H, Fixed, 95% Cl	
Adams3 2018 <sup>19</sup>	6	12	4	12	17.1%	2.00 [0.38, 10.41]		
Emens 2018 <sup>(10)</sup>	13	91	0	21	5.9%	7.39 [0.42, 129.48]		
Schmid 2018 <sup>DR</sup>	15	185	11	266	70.8%	2.05 [0.92, 4.56]		
Sherene 2019 <sup>[28]</sup>	1	40	0	12	6.2%	0.95 [0.04, 24.81]		
Total (95% Cl)		328		311	100.0%	2.28 [1.16, 4.48]	•	
Total events	35		15					
Heterogeneity: Chi <sup>2</sup> = 1	.02, df = 3 (	(P = 0.8	0); I² = 0%				0.001 0.1 1 10	0 1000
Test for overall effect: 2	Z = 2.40 (P	= 0.02)					Favours [PD-L1 negative] Favours [P	

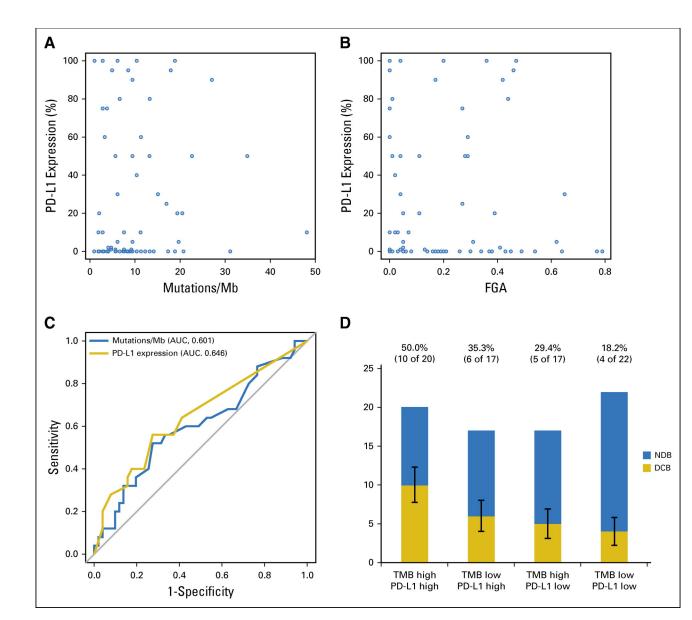
Zou et al. 2020

#### PD-L1 expressed on cancer cells or immune cells?

#### Current FDA Approved Biomarkers for ICIs

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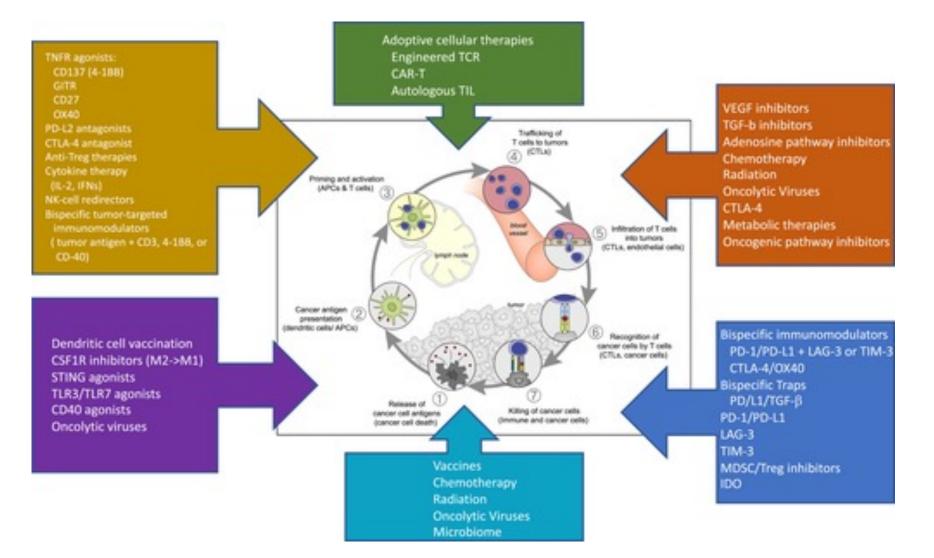
#### Tumor mutational burden (TMB)



NBD = no durable benefit DCB = durable clinical benefit

Rizvi et al. Thoracic Oncology 2021

# Need for better biomarkers: Cancer immunity cycle, revisited



# **Immune Contexture of Tumors**

Table. Comparison of Pooled Diagnostic Sensitivity and Specificity and Predictive Values for Responders vs Nonresponders After Anti-PD-1/PD-L1 Therapy Between Different Assay Modalities<sup>a</sup>

Mod	ality	Pooled Sensitivity	Pooled Specificity	Pooled PPVs	Pooled NPVs
PD-L	.1 IHC (n = 24)	0.50 (0.48-0.53)	0.63 (0.62-0.65)	0.34 (0.32-0.36)	0.78 (0.76-0.79)
TMB	(n = 10)	0.57 (0.51-0.62)	0.70 (0.66-0.73)	0.42 (0.38-0.47)	0.80 (0.77-0.83)
GEP	(n = 9)	0.71 (0.67-0.75)	0.51 (0.48-0.54)	0.42 (0.39-0.46)	0.77 (0.74-0.81)
mIH	C/IF (n = 7)	0.60 (0.53-0.66)	0.78 (0.73-0.82)	0.63 (0.56-0.70)	0.75 (0.70-0.80)
Mult	imodality (n = 6)	0.58 (0.50-0.65)	0.79 (0.75-0.82)	0.41 (0.33-0.48)	0.88 (0.85-0.91)

Abbreviations: GEP, gene expression profiling; mIHC/IF, multiplex immunohistochemistry/ immunofluorescence; PD-L1 IHC, programmed cell death ligand 1 immunohistochemistry; TMB, tumor mutational burden.

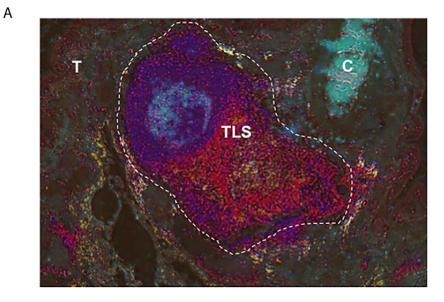
<sup>a</sup> All data are reported as a proportion (95% Cl). Nonoverlapping 95% Cls suggest statistical significance.



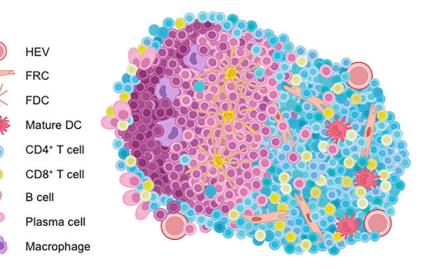
From: Comparison of Biomarker Modalities for Predicting Response to PD-1/PD-L1 Checkpoint Blockade: A Systematic Review and Meta-analysis

JAMA Oncol. Published online July 18, 2019. doi:10.1001/jamaoncol.2019.1549

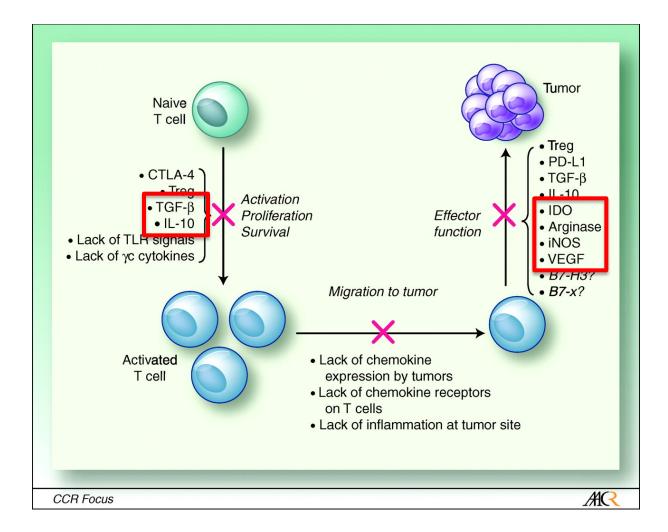
### Tertiary Lymphoid Structures (TLS)



В



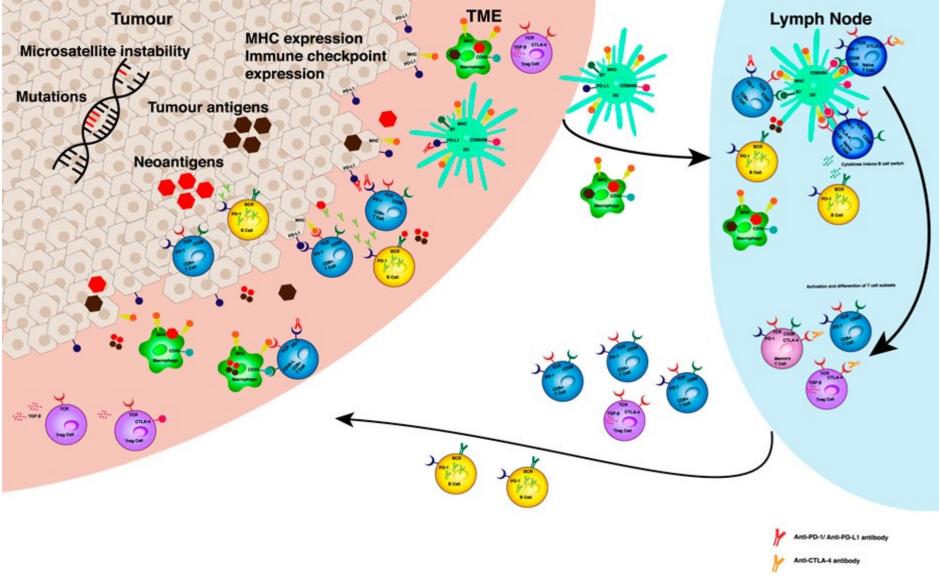
### Many barriers in vivo to effective antitumor T cell immunity



Lizée G et al. Clin Cancer Res 2007;13:5250-5255

# Moving beyond the tumor

### **Tumor-Draining Lymph Nodes**



Goode et al. Frontiers in Molecular Biosciences 2021

### **Blood Biomarkers**

- Immune cell subsets: PD1+ CD8 T cells, Tregs
- Serum cytokines: IL-6
- Immune cells signaling responses
- T cell receptor (TCR) clonality
- Peripheral blood shed PD-L1
- Cell-free (cf)DNA

# Multi-parameter blood biomarkers

Biomarker	Cancer type	N	No. of patients	Main results	Reference
(%Ki-67 <sup>+</sup> cells/PD-1 <sup>+</sup> CD8 <sup>+</sup> T cells 3-wk post- treatment)/baseline tumor burden (Ki67/TB)	Melanoma	Discovery cohort: 23		Higher Ki67/TB significantly associated with superior ORR (p=0.03) and PFS (p=0.004).	Huang et al. (34)
		Validation cohort: 18		Higher Ki67/TB associated with superior ORR (p=0.14) and PFS (p=0.06).	
(%Ki-67 <sup>+</sup> cells/PD-1 <sup>+</sup> CD8 <sup>+</sup> T cells 1-wk post- treatment)/(%Ki-67 <sup>+</sup> cells/PD-1 <sup>+</sup> CD8 <sup>+</sup> T cells a baseline) (Ki-67 <sup>D7/D0</sup> )		Discovery cohort: 31		Higher Ki-67 <sup>D7/D0</sup> significantly associated with durable clinical benefit (PR, or SD for 6 months or longer; p<0.001) and PFS (p=0.027)	Kim et al. (32)
	NSCLC	Discovery cohort: 33		Higher Ki-67 <sup>D7/D0</sup> significantly associated with durable clinical benefit (PR, or SD for 6 months or longer; p<0.01), PFS (p=0.004), and OS (p=0.001)	
		Validation cohort: 46		Higher Ki-67 <sup>D7/D0</sup> significantly associated with durable clinical benefit (PR, or SD for 6 months or longer; p<0.01), PFS (p=0.002), and OS (p=0.037)	
%FoxP3 <sup>¬</sup> PD-1 <sup>hi</sup> CD4 <sup>+</sup> T cells/CD4 <sup>+</sup> T cells (4PD1 <sup>hi</sup> ) 3-wk post-treatment Fold change of 4PD1 <sup>hi</sup>	Melanoma	52		Higher frequency of $4PD1^{hi}$ 3-wks post treatment (p=0.0005) and fold change of $4PD1^{hi}$ (p=0.046) associated with poorer OS.	Zappasodi et al. (36)
$\Gamma CR$ diversity of PD-1 <sup>+</sup> CD8 <sup>+</sup> T cells at baseline and post-treatment	NSCLC	Discovery cohort: 25 Validation cohort: 15		Higher baseline diversity in PD-1 <sup>+</sup> CD8 <sup>+</sup> T cells ( $p=0.021$ ) and increased clonality after treatment ( $p=0.002$ ) associated with superior PFS.	Han et al. (39)
%CD27 <sup>-</sup> CD28 <sup>-</sup> cells/CD4 <sup>+</sup> T cells at baseline	NSCLC	51		Higher frequency of CD27 <sup>-</sup> CD28 <sup>-</sup> CD4 <sup>+</sup> T cells associated superior PFS (p=0.001).	Zuazo et al. (42)
Ratio of the frequency of Treg cells and PMN- MDSCs at baseline	NSCLC	Discovery cohort: 34		Higher ratio of the frequency of Treg cells and PMN-MDSCs associated with superior PFS (p=0.0079).	Kim et al. (44)
		Validation cohort: 29		Higher ratio of the frequency of Treg cells and PMN-MDSCs associated with superior PFS (p=0.0017).	
%Effector/memory (CCR7 <sup>-</sup> CD45RA <sup>-</sup> ) cells/CD8 <sup>+</sup> T cells at baseline	NSCLC	263 (flow cytometry ana	lysis in 144)	Lower frequency of effector/memory $\text{CD8}^+$ T cells with development of hyperprogressive disease (p<0.001) and poor PFS (p<0.001) and OS (p<0.001).	Kim et al. (53)
%TIGIT <sup>+</sup> cells/PD-1 <sup>+</sup> CD8 <sup>+</sup> T cells at baseline				Higher frequency of TIGIT <sup>+</sup> cells among PD-1 <sup>+</sup> CD8 <sup>+</sup> T cells in peripheral blood at baseline significantly associated with development of hyperprogressive disease ( $p<0.001$ ) and poor PFS ( $p<0.001$ ) and OS ( $p=0.01$ ).	

NSCLC, non-small-cell lung cancer; ORR, objective response rate; OS, overall survival; PFS, progression-free survival; PR, partial response; SD, stable disease; TET, thymic epithelial tumor; CCR7, C-C chemokine receptor type 7.

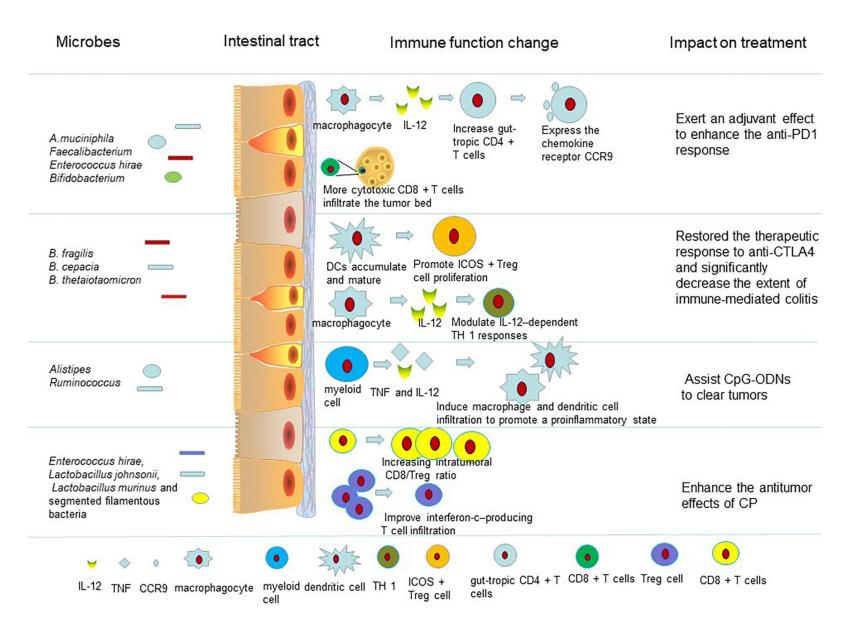
### Functional (dynamic) immune assays

- Most assays measure quantities (static)
- Immune system is dynamic -> need to assess functional status of patients' immune system
- Measure functional responses after specific interrogation:
  - Cytokine signaling responses
  - Cytotoxicity responses
  - Cytokine production

# **Biomarkers for irAEs**

Biomarker	Cancer type	# pts	Main results	Reference
Fold change of effector Treg cells 1-wk post-	TET	31	Patients with irAEs can be distinguished into 4 distinct	Kim et al.
Th17 to Th1 ratio at baseline	NSCLC	60	subtypes according to the T-cell parameters and each	(63)
%Ki-67 <sup>+</sup> /PD-1 <sup>+</sup> CD8 <sup>+</sup> T cells 1-wk post-treatment			T-cell parameter predicts the corresponding subtype of	
%TNF- $\alpha^+$ /CD4 <sup>+</sup> or CD8 <sup>+</sup> T cells 1-wk post-treatment			irAEs	
Memory cytotoxic (CD45RO <sup>+</sup> GzmB <sup>+</sup> Ki-67 <sup>+</sup> ) CD4 <sup>+</sup> T	Melanoma	3	Activated memory CD4 <sup>+</sup> T cells were highly enriched	Johnson et al.
cells			in inflammed, affected region of cases with	(68)
Early B cell changes (decline in B cells, increase in	Melanoma	23	Decline in B cells but an increase in CD21 <sup>10</sup> B cells	Das et al.
CD21 <sup>lo</sup> B cells)			more prominent in patients with severe irAEs that	(72)
			received combined anti-PD-1 and anti-CTLA-4	
Cytokine expression-based score	Melanoma	98	Eleven cytokines were integrated into a single score	Lim et al.
			(CYTOX) and it significantly predicted development of	(77)
			severe irAEs in patients treated with combined anti-PD-	
		49	CYTOX score significantly predicted development of	
Auto-Abs (rheumatoid factor, antinuclear Ab,	NSCLC	137	Preexisting rheumatoid factor or auto-Abs significantly	Toi et al. (75)
antithyroglobulin, and antithyroid peroxidase)			correlates with development of any grade irAEs	
Anti-thyroid Abs (anti-microsomal and anti-	NSCLC	51	Presence of anti-thyroid Abs either at baseline or	Osorio et al.
thyroglobulin)			during anti-PD-1 treatment was significantly associated	(76)
NSCLC, non-small-cell lung cancer; TET, thymic epith	elial tumor.			

# **GI** Microbiome



#### Current state of biomarkers: Work in progress

	Biomarker			Method of detection	T	issue type	Association with positive predictive value	Level of evidence' and result
umon-related	PD-L1 pathway	athway Tissue PD-L1 Soluble PD-L1		Immunohistochemistry		umor	Positive PD-L1 tumor expression	Concordant results
Antigen rec				ELISA chemituminescence		bood	Conflicting results	Conflicting results
	Antigen recognitio	on MSI-MDF	0	Immunohistochemistry		umor	Mismatch repair deficiency: hypermutator phenotypes	Concordant results
		TMB	Turnoral TMB	WES; NGS (Foundation One CDx <sup>16</sup> and MSF	K-IMPACT) T	umor	High mutational rate (ITMBa175 mut/exome)	Conflicting results
			Blood TMB	WES; NGS	в	bood	High mutational rate	Conflicting results
		Tumor-sp	ecific genotype	Direct sequencing; NGS	Ti.	umor	Absence of EGFR, ALK or ROS1 mutations	Conflicting results
	Others	CTC		Enrichment (CellSearch*) and detection (IF a	staining) B	Rood	Low baseline CTC count	Results to be validated
		ctONA		Multiplex targeted NGS: digital droplet PCR: SNP array	RNASeq: B	lood	Undetectable ctDNA levels at week 8 of treatment; early decrease in ctDNA burden at 1 month of treatment	Concordant results
		Epigenetic		Bisuffite conversion of genomic DNA, whole-genome amplification and array-based capture and scoring of CpG loci		umar	EPIMMUNE signature	Results to be validated
umon/host interaction:			Immunohistochemistry; flow cytometry		umor	CD3 CD8' infitration; TILs density-5%	Concordant results	
nicroenvironment			Immunohistochemistry; flow cytometry		umor	TLS presence	Results to be validated	
lost-related	Gene expression	signature		Microarray analysis	T.	umor	IFN-y gene signature	Concordant results
	Circulating immur cells	Circulating immune CD3 <sup>+</sup> T cells cells		Complete blood count; flow cytometry		bood	Expansion of PD-1° CD8 T cells during treatment; low baseline proportion of CD28 CD57°KLPG1° CD8 T cells	Results to be validated
		Neutrophils and MDSC		Complete blood count; flow cytometry		Rood	Low baseline proportion of circulating M-MDSCs; low baseline dNLR	Results to be validated
	Soluble systemic	ine or cap		Spectrophotometry		lood	Low baseline LDH levels	Conflicting results
	immune or inflammatory			Immunoturbidimetry		lood	Low baseline CRP levels	Results to be validated
	markers	Abumin		Immunoturbidimetry	8	lood	High baseline albumin levels	Results to be validated
Microbiota		Cytokines		EUSA chemiluminescence		liood	Early decrease of 8.–8 during treatment; increase of TNF-s and IFN-y during treatment	Results to be validated
	Microbiota			Bacterial 165 ribosomal RNA gene sequencing		iut microbiota	High gut diversity at baseline Akkermansia enrichment	Results to be validated
	Prospective study 1			Cono	Concordant results A			
	B	Introspective	study with indepe	ndent cohorts 2 Confi	ficting results B	1		

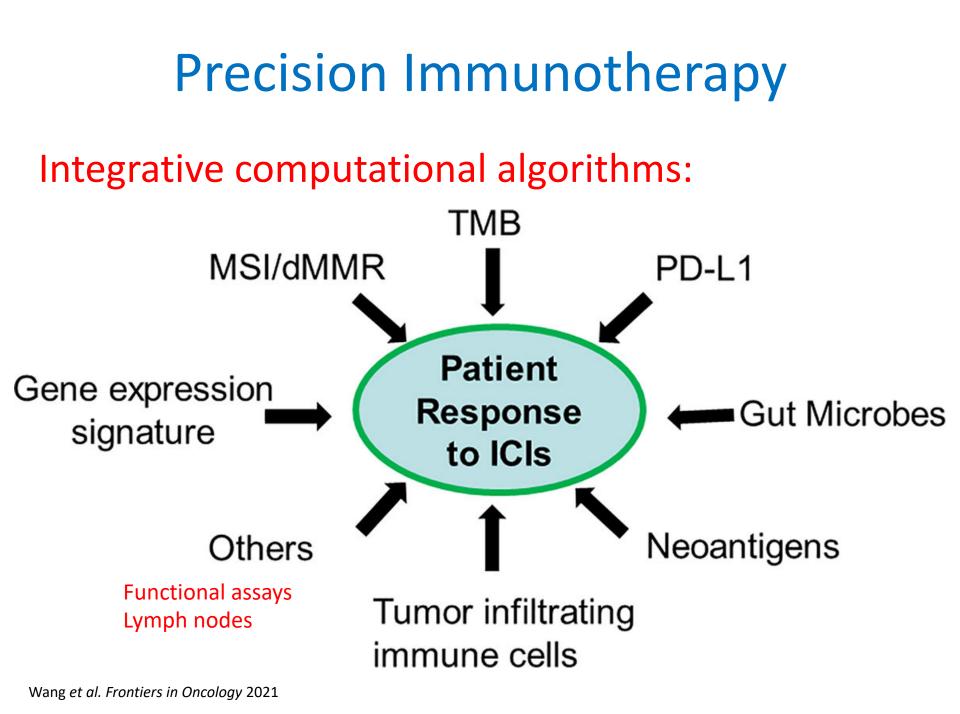
Retrospective study without a control group 3 Results to be validated in further studies C

Duchemann et al. Translational Lung Cancer Research 2021

# Biomarkers for CAR T therapy?

IFNγ production by CAR T cells and IFNγ responsiveness of host immune cells are critical for tumor immune landscape remodeling to promote a more activated and less suppressive tumor microenvironment.

Alizadeh et al. Cancer Discovery 2021



# **Thanks!**

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