

How Liquid Biopsy Technology is Delivering Personalized Therapy in NSCLC

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Professor in Medicine

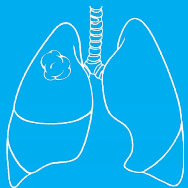
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The Tisch Cancer Institute

Mount Sinai, New York, NY, US



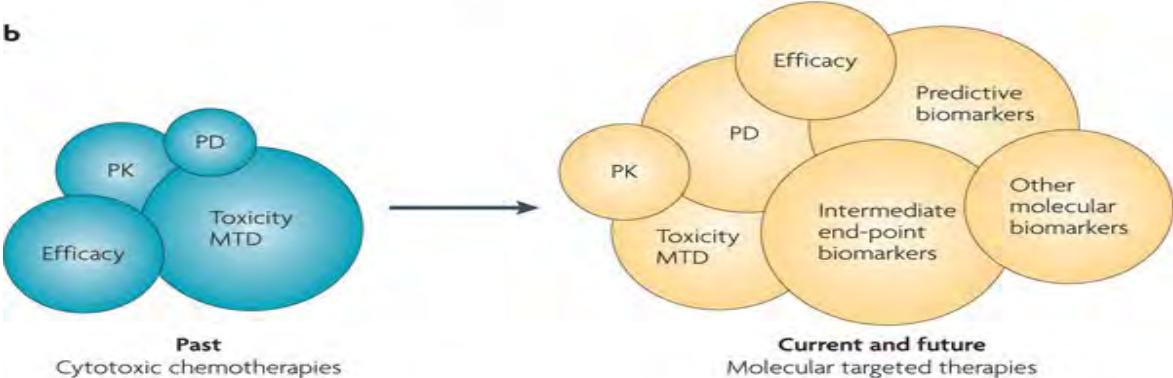
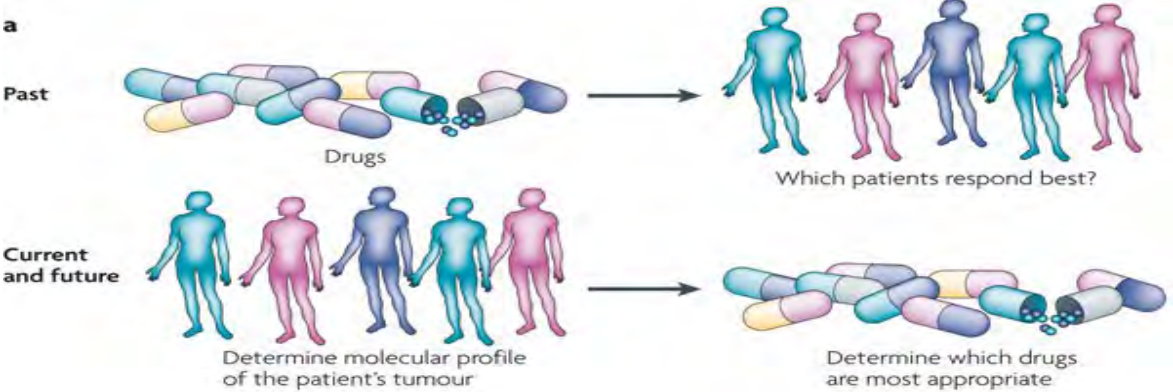
Center for Thoracic Oncology



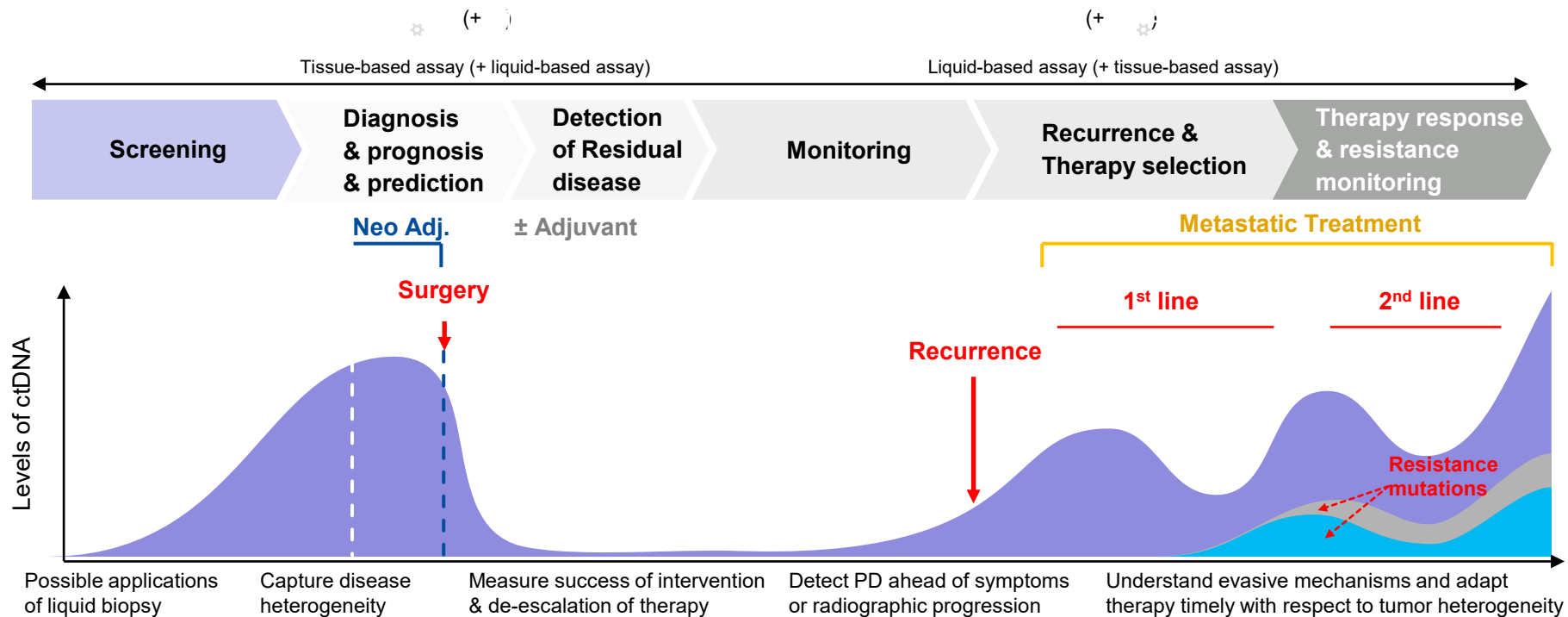
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Current Treatment and Trial Paradigm



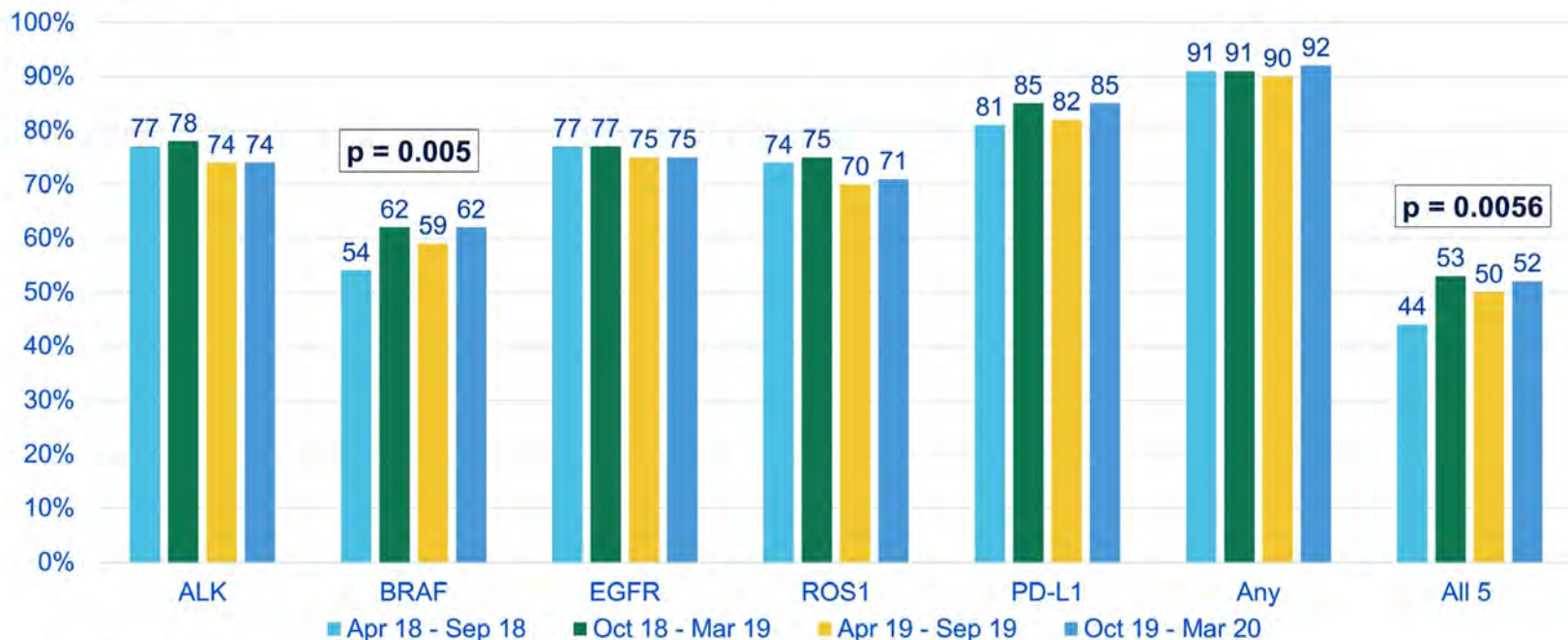
Liquid biopsy can provide clinically-valuable information along the whole patient journey



PD: progressive disease.

Adapted from Wan, J.C.M., et al., (2017) *Nat Rev Cancer* 17:223-

Biomarker testing rates over time for overall study population



p values: Cochran Armitage test for trend analysis

Presented By: **Nicholas J. Robert, MD**
On behalf of the MYLUNG Consortium™

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NGS testing rates over time for the overall population



p value: Cochran Armitage test for trend analysis

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Tissue biopsy may not capture the genomic landscape of a patient's entire tumour burden

Intratumour heterogeneity



The genomic landscape **within a single tumour manifestation** may not be uniform

Tissue biopsy may not capture subclonal populations of tumour cells with distinct alterations

Intrapatient heterogeneity

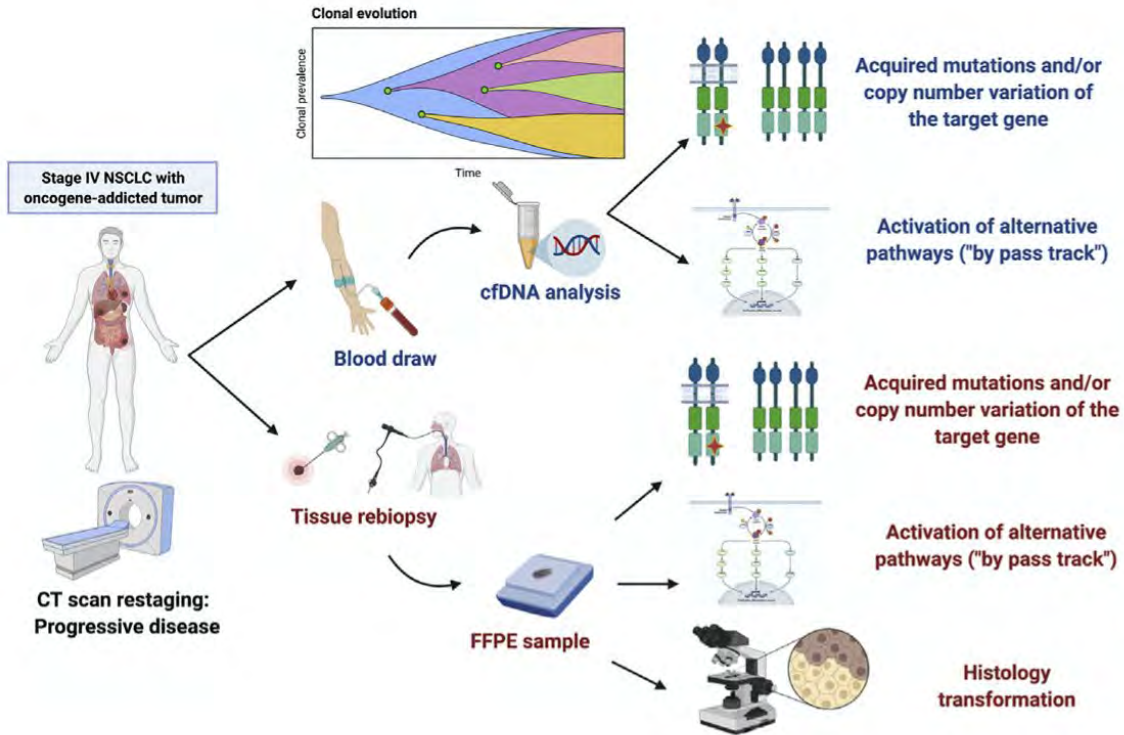


The genomic landscape may **differ between tumour sites** within a patient

Tissue biopsy from a single lesion will miss alterations unique to other lesions

**As well as spatial heterogeneity, as the genomic landscape of a cancer evolves over time, temporal heterogeneity should also be considered
Therefore archival tissue may not fully represent the tumour genotype at progression**

Liquid biopsy and tissue rebiopsy after acquired resistance to TKI



Main liquid biopsy techniques used

NGS-based approaches:

- ✓ High sensitivity
- ✓ Multiplex
- ✓ Gene rearrangements
- ✓ Gene amplifications

PCR-based approaches:

- ✓ Variable sensitivity
- ✓ Single gene testing
- ✓ Only for mutations

Main techniques used for tumor tissue

NGS-based approaches:

- ✓ High sensitivity
- ✓ Multiplex
- ✓ Gene rearrangements
- ✓ Gene amplifications

FISH:

- ✓ Gene rearrangements & amplifications

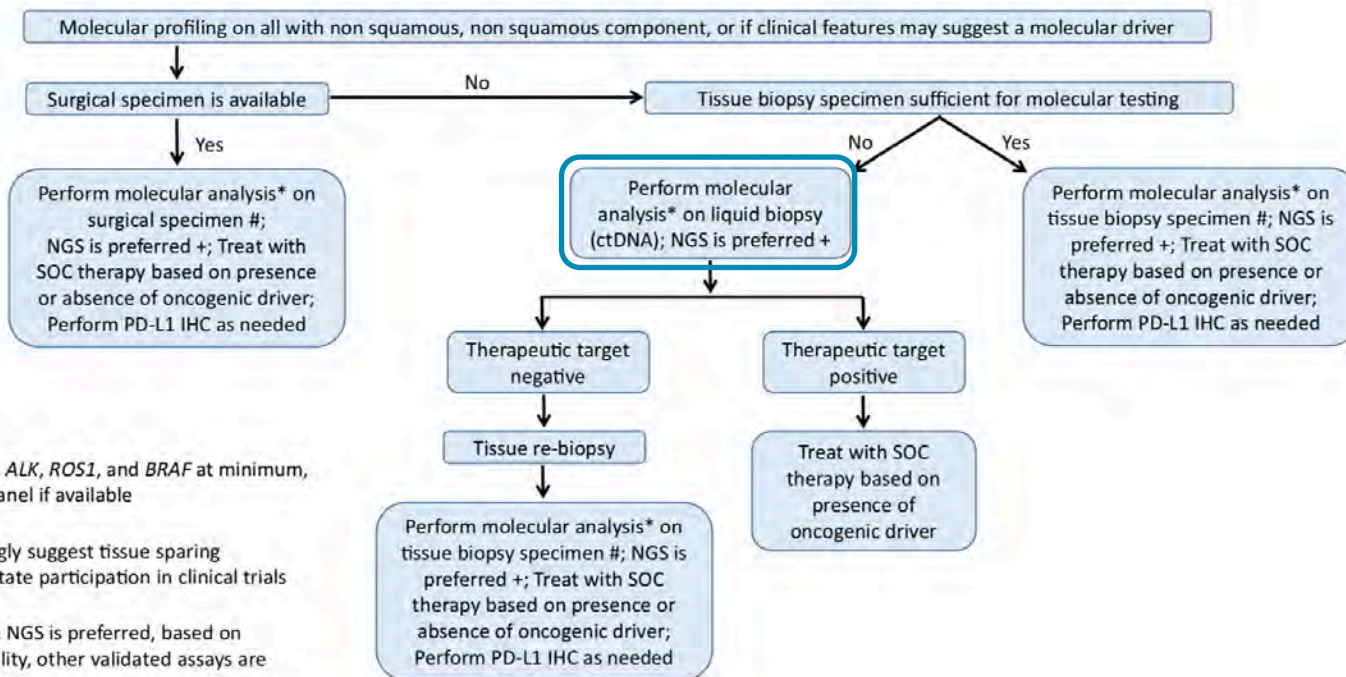
PCR-based approaches:

- ✓ Variable sensitivity
- ✓ Single/Multiplex gene testing
- ✓ Only for mutations

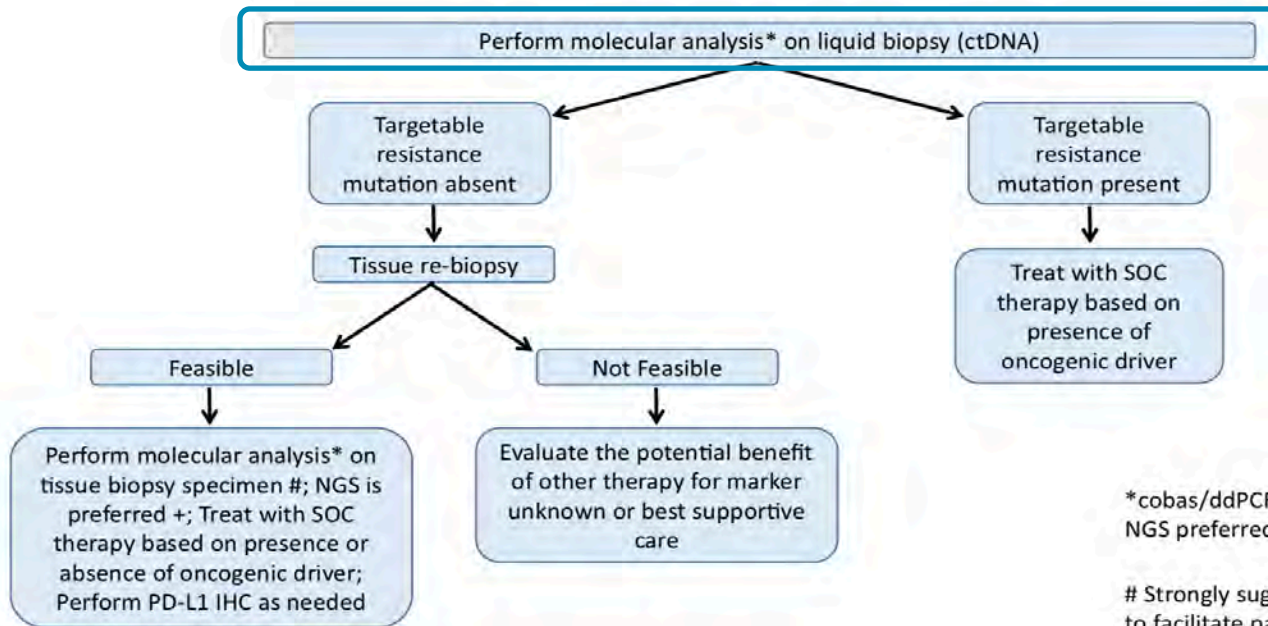
IHC:

- ✓ Protein expression

Patients with advanced treatment-naive NSCLC



Patients with progressive or recurrent NSCLC during treatment with TKI



*cobas/ddPCR for *EGFR* mutation
NGS preferred for *ALK* and *ROS1*

Strongly suggest tissue sparing
to facilitate participation in clinical trials

+ While NGS is preferred, based on
availability, other validated assays are
acceptable

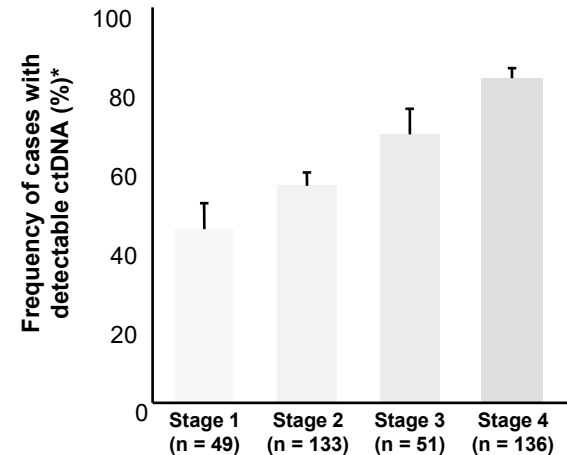
Analysis of circulating tumour DNA (ctDNA) poses distinct challenges



ctDNA

- constitutes a **highly variable fraction** of the total plasma cfDNA **from < 0.1% to > 90%**^{1,2}
 - if ctDNA fraction is low, detection of **alterations is more challenging**^{2,3}
 - need to be able to **detect mutations down to $\leq 0.1\%$ MAF** (particularly for detection of MRD)^{3,4}
- is more **fragmented at 134 - 144 bp**, compared with ~ 166 bp fragments of 'normal' plasma cfDNA⁵
- has a very short **half-life of less than one hour** in circulation^{2,6}

Amount of shedded, or detectable, ctDNA is variable depending on factors such as tumour stage, histology, vascularity and treatment^{1,5-8}



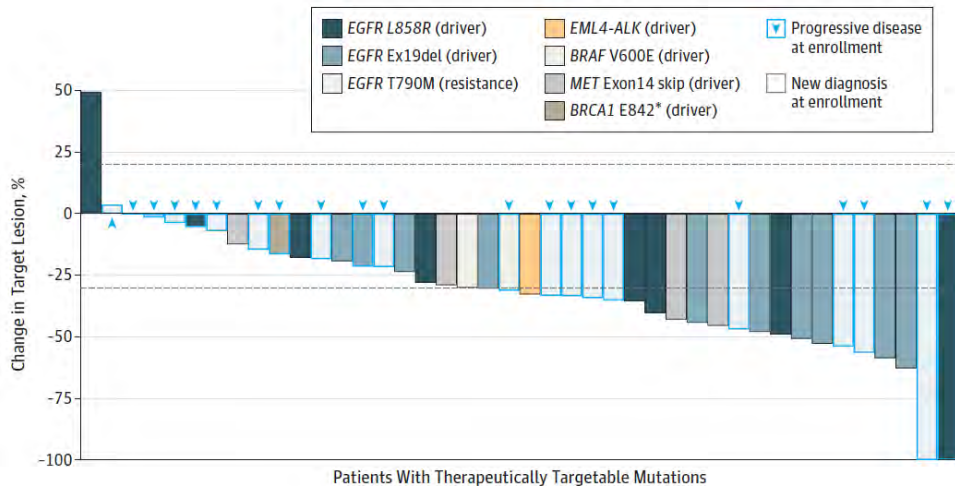
Somatic cfDNA alterations were detected in 85% (18,503 / 21,807) of patients across various cancer types⁹

* Figure adapted from reference 5. cfDNA: cell-free DNA; ctDNA: circulating tumour DNA; MAF: mutant allele frequency; MRD: minimal residual disease.

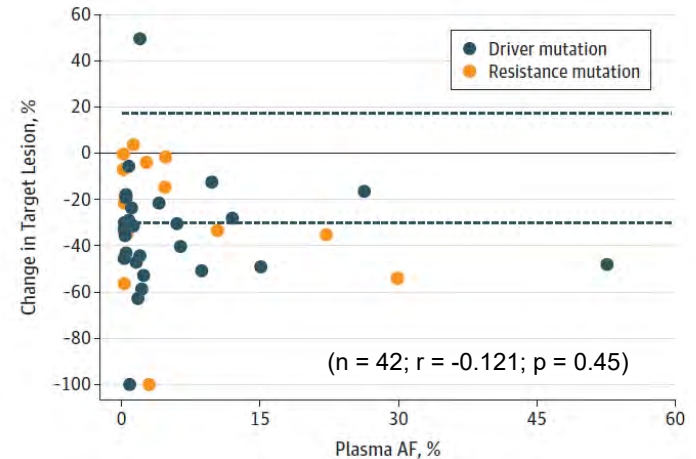
1. Hinrichsen, T., et al. (2016) *J Lab Med* 40:313-22; 2. Corcoran, R.B. and Chabner, B.A. (2018) *N Engl J Med* 379:1754-65; 3. Johansson, G., et al. (2019) *Biomol Detect Quantif* 17:100078; 4. Jennings, L. et al. (2017) *J Mol Diagn* 19:341-65 5. Wan, J.C.M., et al., (2017) *Nat Rev Cancer* 17:223-38; 6. Maltoux, A. K., et al (2019) *Sci Transl Med* 11:eaay1984; 7. Bettegowda, C., et al. (2014) *Sci Transl Med* 6:224ra24; 8. Diaz, L.A. and Bardelli, A. (2014) *J Clin Oncol* 32:579-86; 9. Zill, O.A., et al. (2018) *Clin Cancer Res* 24:3528-38.

Plasma-based biomarkers with low allele frequency may still respond to targeted therapy

Responses to plasma-indicated targeted therapy by RECIST



Correlation of RECIST and allele frequency



AF: allele frequency; RECIST: Response Evaluation Criteria in Solid Tumours.

Aggarwal, C., et al. (2019) *JAMA Oncol* 5:173-80.

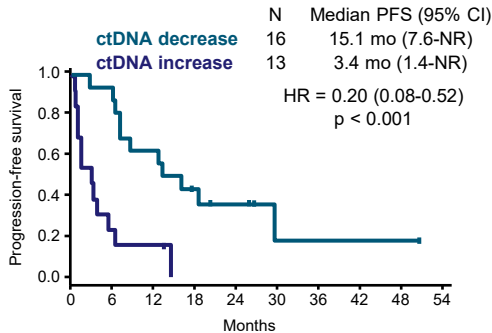
ctDNA kinetics as predictive marker for treatment response or resistance



Identification of early plasma ctDNA changes to predict response to first-line pembrolizumab +/- chemotherapy in aNSCLC patients¹

Blood samples were collected on 1st day of treatment and at each subsequent cycle

A 36-gene panel NGS* detected early quantitative changes across a wide range of variants



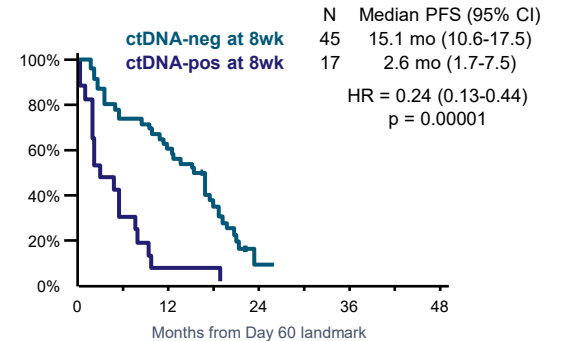
Rapid decrease of ctDNA correlated with clinical benefit, while increase correlated with PD



Residual ctDNA to predict PFS and OS in EGFRmut NSCLC patients treated with afatinib +/- cetuximab²

Blood samples were collected at baseline, cycle 3-day 1 and at progression

A 73-gene panel NGS† detected quantitative changes in EGFRmut ctDNA (primary activating mutations E19del or L858R)



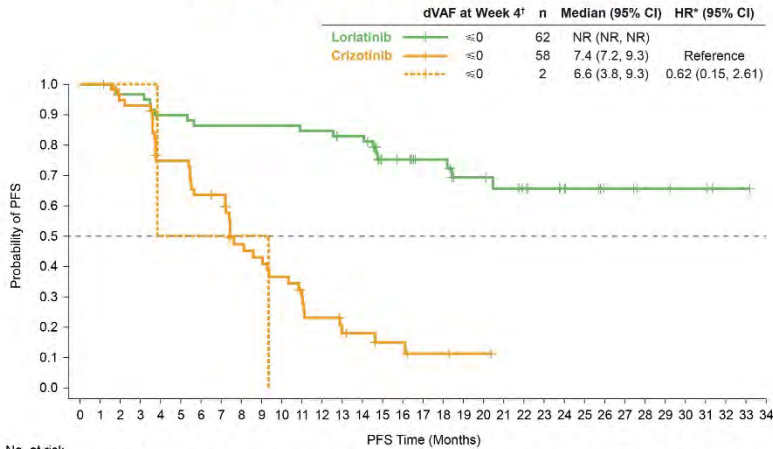
Clearance of EGFR ctDNA after 60 days of therapy correlated with substantial improvement in PFS and OS

In both studies PFS is significantly longer in NSCLC patients with early ctDNA decrease / clearance
These results suggest a potential role for ctDNA NGS analysis to detect pharmacodynamic biomarkers of response or resistance to targeted therapies and immunotherapies

*Samples were analysed in the Inivata CLIA-accredited laboratory (Research Triangle Park, NC) for InVision ctDNA analysis. †Tested by Guardant Health, Inc. using G360 panel. aNSCLC: advanced non-small cell lung cancer; ctDNA: circulating tumour DNA; mo: months; NGS: next-generation sequencing; OS: overall survival; PD: progressive disease; PFS: progression-free survival; pts: patients; wk: weeks.
1. Ricciuti, P.C., et al. (2020) ASCO poster 3518; 2. Mack, P.C., et al. (2020) ASCO poster 9532.

Results: Reduction in dVAF at Week 4 Was Associated With Longer PFS With Lorlatinib

ALK Fusion and/or Mutation

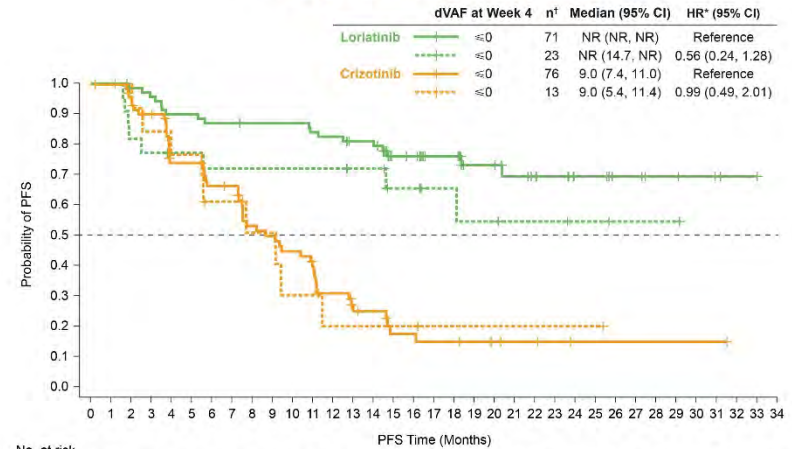


	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34		
Lorlatinib dVAF ≤0:	62	62	57	57	52	52	50	50	50	50	50	49	49	46	46	32	31	26	26	20	20	18	16	13	11	9	6	6	4	4	3	3	1	1	0	0	
Crizotinib dVAF ≤0:	58	58	53	52	40	40	34	33	22	20	17	12	10	7	6	4	4	2	2	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Crizotinib dVAF >0:	2	2	2	2	1	1	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

*HR <1.00 favors dVAF ≤0.

[†]No patients in the lorlatinib group had a dVAF >0 (ALK fusion and/or mutation) at week 4.
HR, hazard ratio; NR, not reached.

Any Somatic Alteration



	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34		
Lorlatinib dVAF ≤0:	71	71	68	66	62	62	60	60	59	59	59	57	56	51	38	37	30	30	22	22	19	17	13	11	9	6	6	4	4	3	3	1	1	0	0	0	0
Lorlatinib dVAF >0:	23	23	18	17	15	15	14	14	14	14	14	14	12	12	9	9	6	6	5	3	3	2	2	1	1	1	1	1	1	1	1	0	0	0	0	0	0
Crizotinib dVAF ≤0:	76	73	65	63	50	50	44	43	32	30	27	21	17	12	11	7	7	6	5	4	3	3	2	1	1	1	1	1	1	1	1	1	1	0	0	0	0
Crizotinib dVAF >0:	13	13	12	11	10	10	7	7	5	5	3	3	2	2	2	2	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0

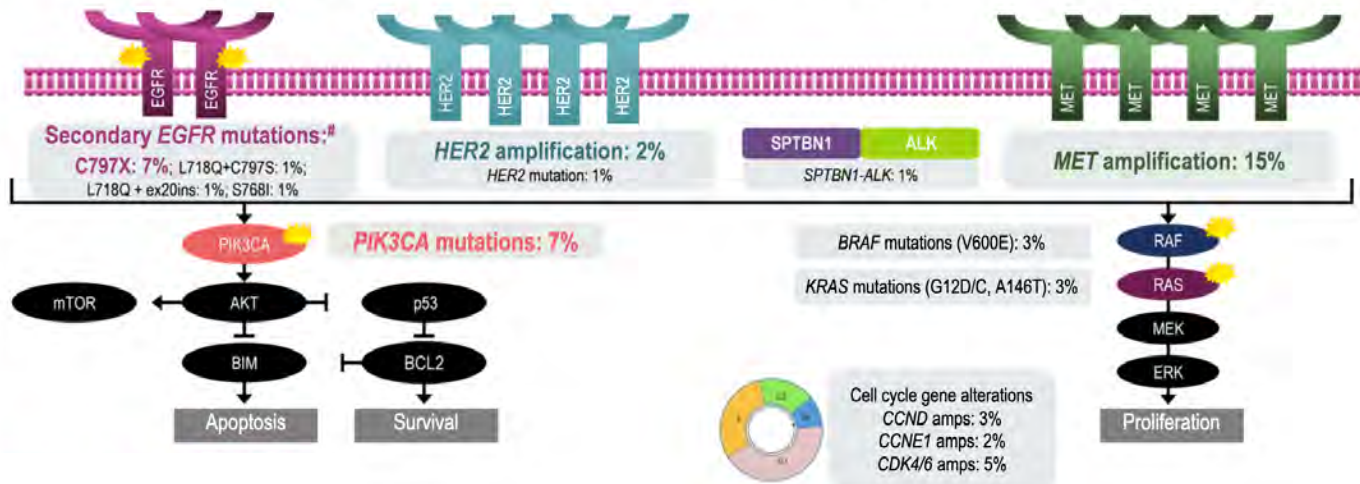
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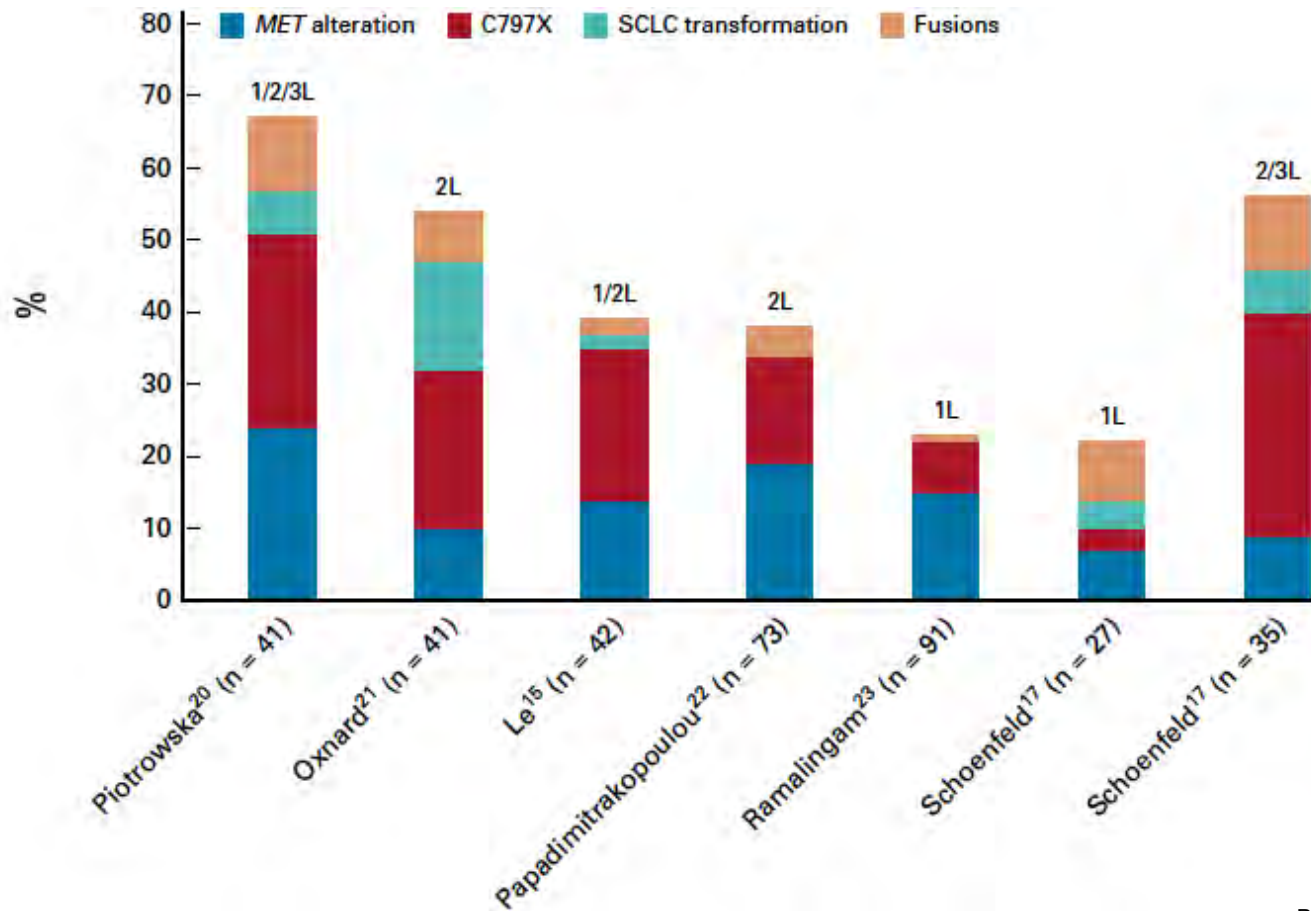
Acquired mechanisms of resistance to 1st line Osimertinib: The FLAURA analysis (LB)

- No evidence of acquired EGFR T790M
- The most common resistance mechanisms were *MET* amplification and EGFR C797S mutation
 - Other mechanisms included *HER2* amplification, *PIK3CA* and *RAS* mutations

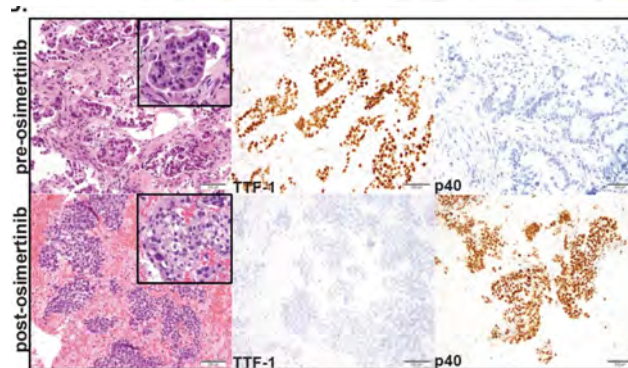
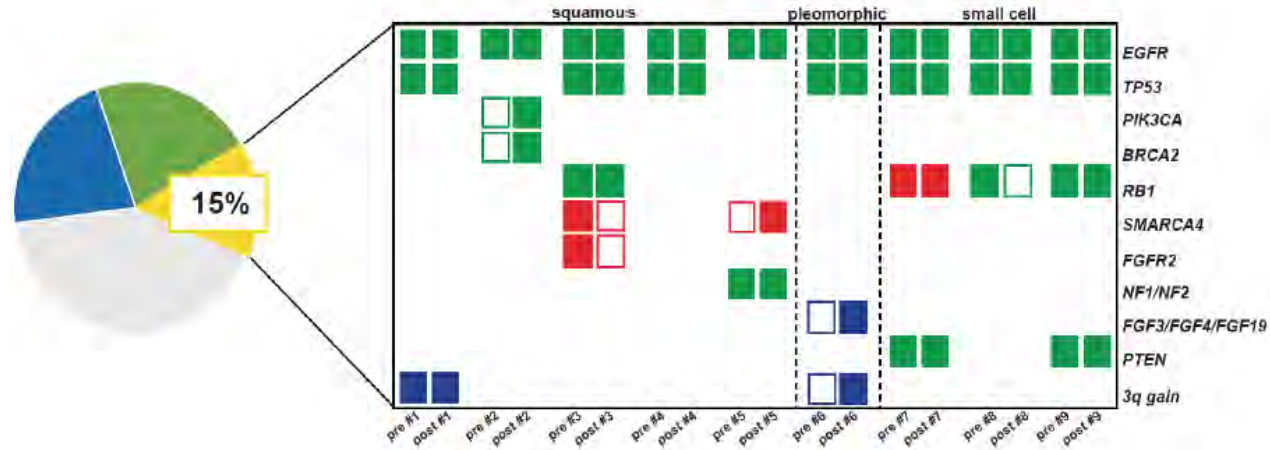


[#]Resistance mechanism reported may overlap with another; [#]Two patients had *de novo* T790M mutations at baseline of whom one acquired C797S at progression

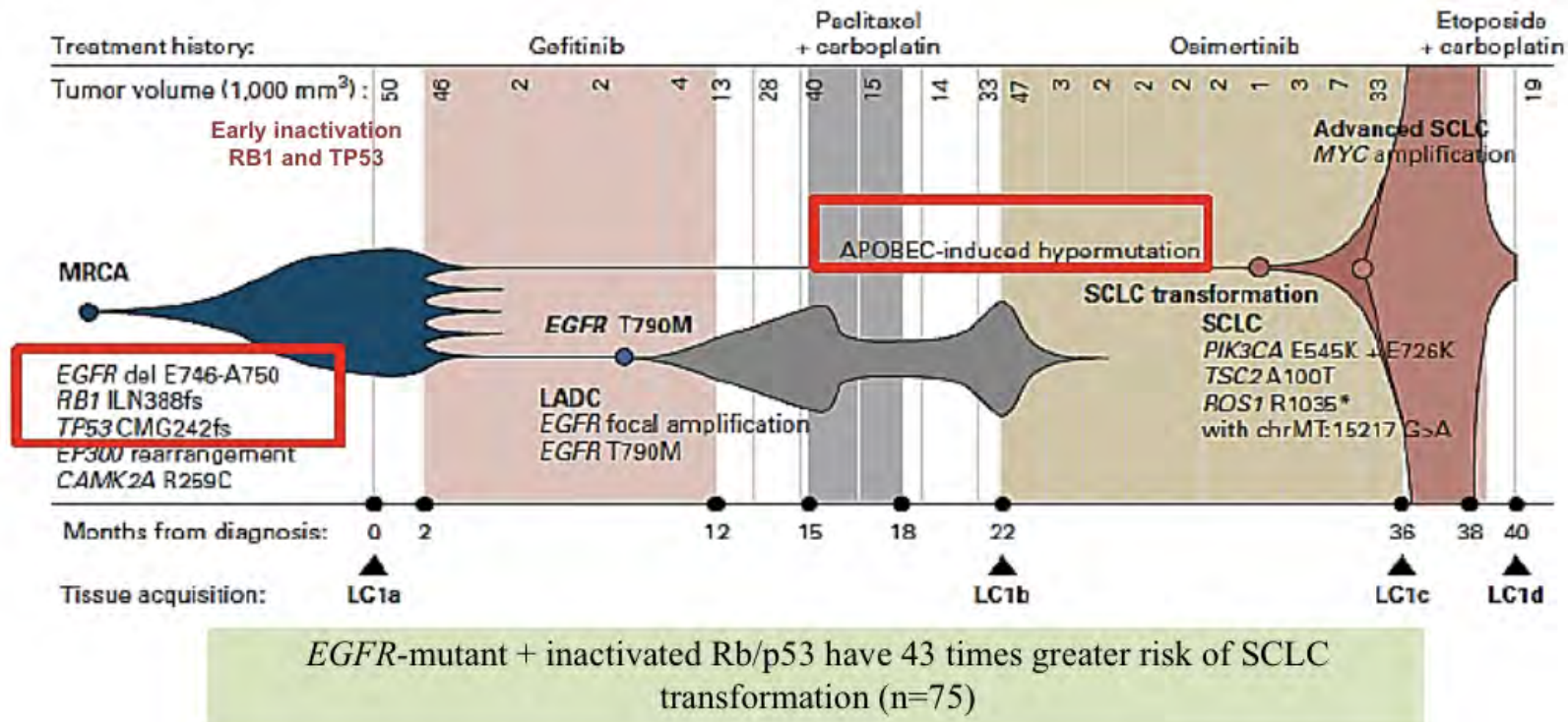
Osimertinib resistance series



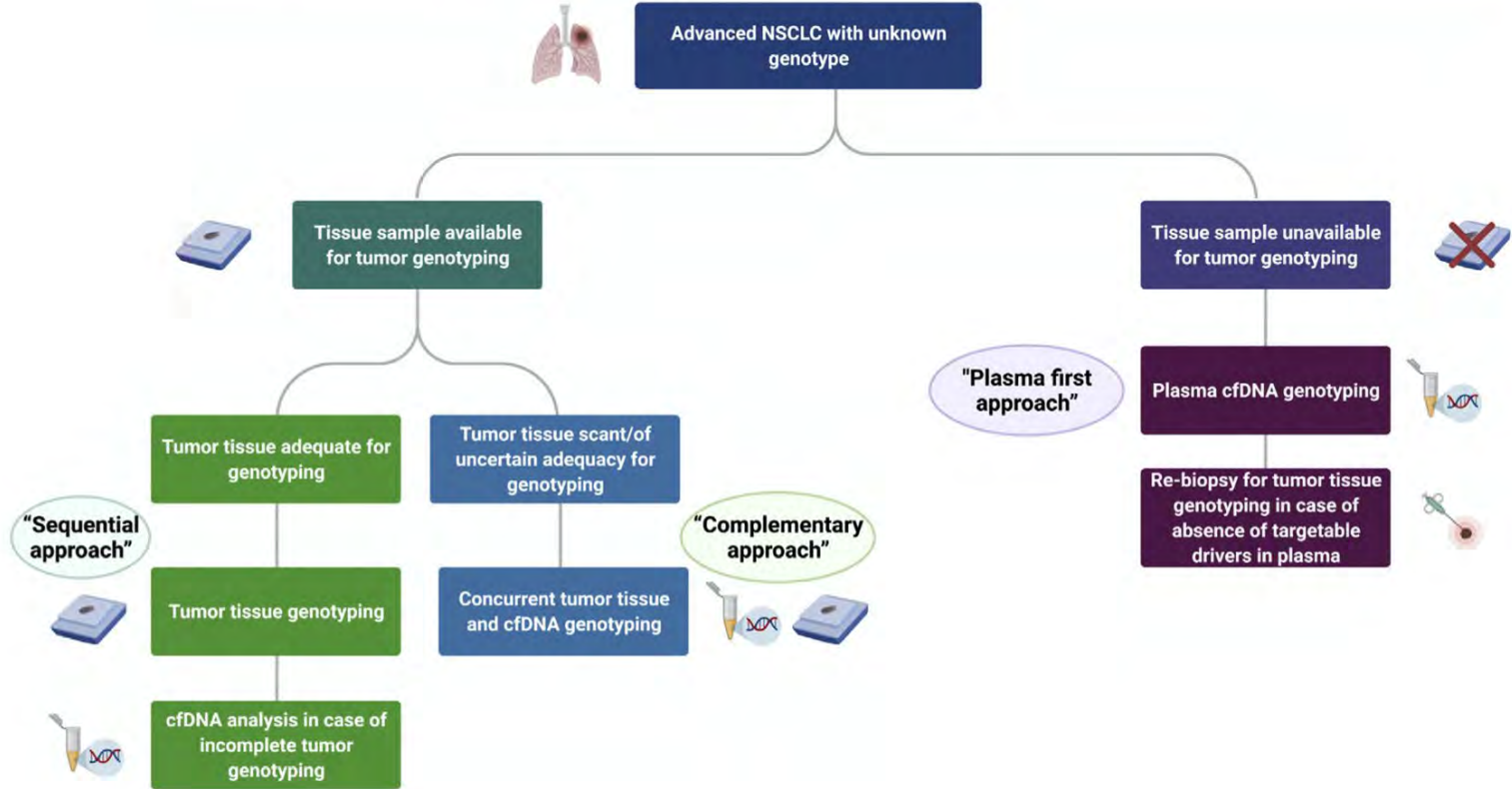
The genomic profile of patients with histologic transformation subtype

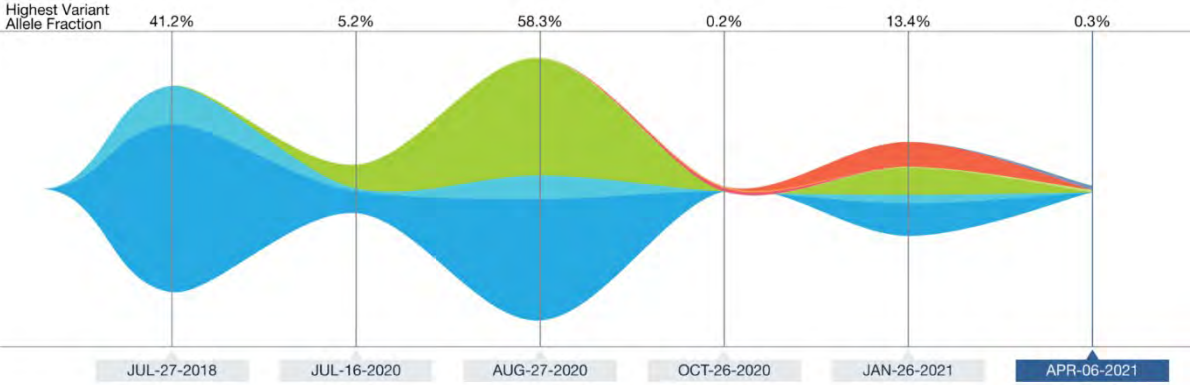


Early Clonal loss of Rb1 and Tp53 predict SCLC transformation

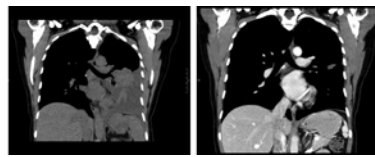


Diagnostic algorithm for liquid biopsy use in treatment-naive advanced/metastatic NSCLC





EGFR Amplification	ND	3.4% ND 0.2% ND ND ND ND
EGFR C797S	ND	ND 5.5% 55.0% ND 10.7% ND
EGFR E746_A750del (Exon 19 deletion)	ND	41.2% 1.7% 58.3% ND 13.4% ND
NTRK2 L689L	ND	0.2% ND ND
EGFR N338N	ND	ND ND ND 0.1% ND ND
TP53 S127F	ND	0.07% 0.4% ND 1.6% ND 2.0%
EGFR T790M	ND	ND ND ND ND 9.8% ND

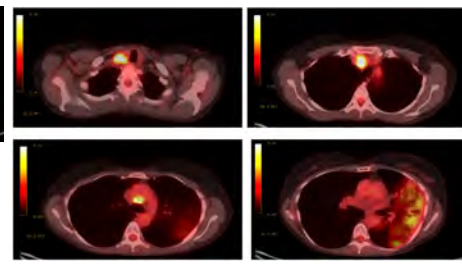


Osimertinib

PFS 24 months

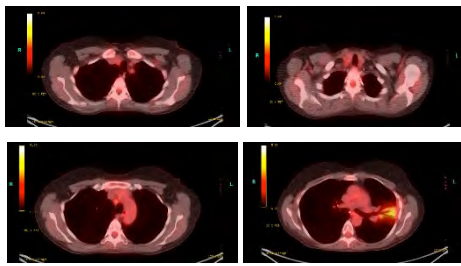


Radiotherapy



Erlotinib

PR 2 months



PFS 5 months



Multiple nodules

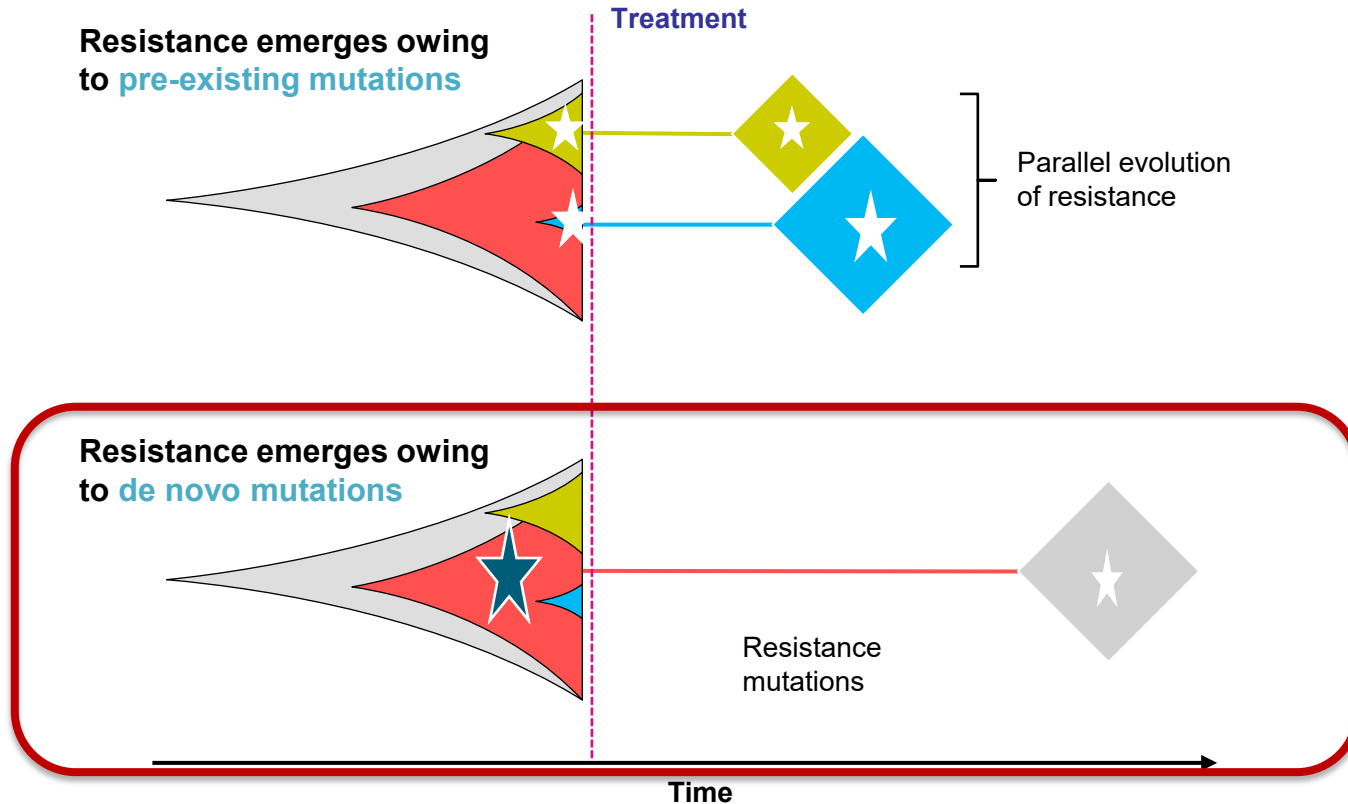


Chemotherapy

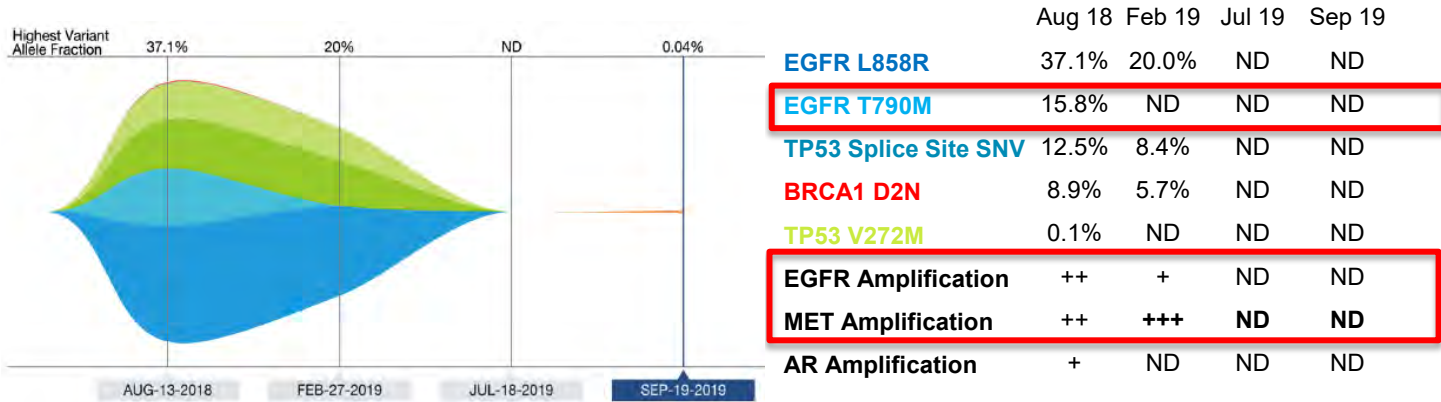


Complete Response

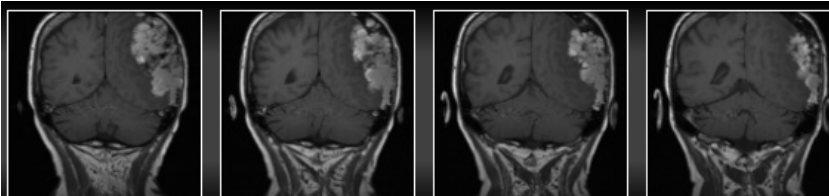
Clonal evolution of treatment resistance



Case #2: 71 year old NSCLC patient



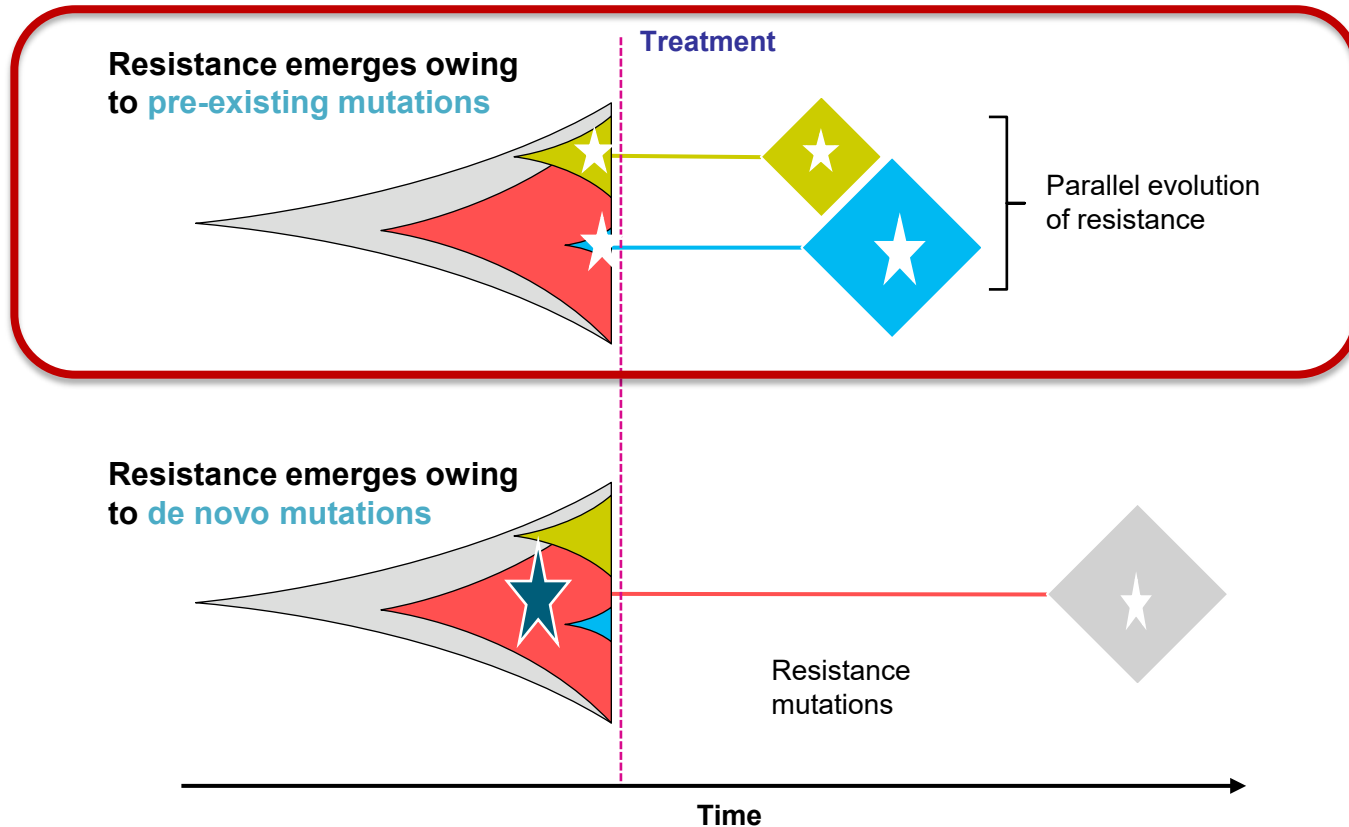
osimertinib start → PFS 6 months → osimertinib + crizotinib start



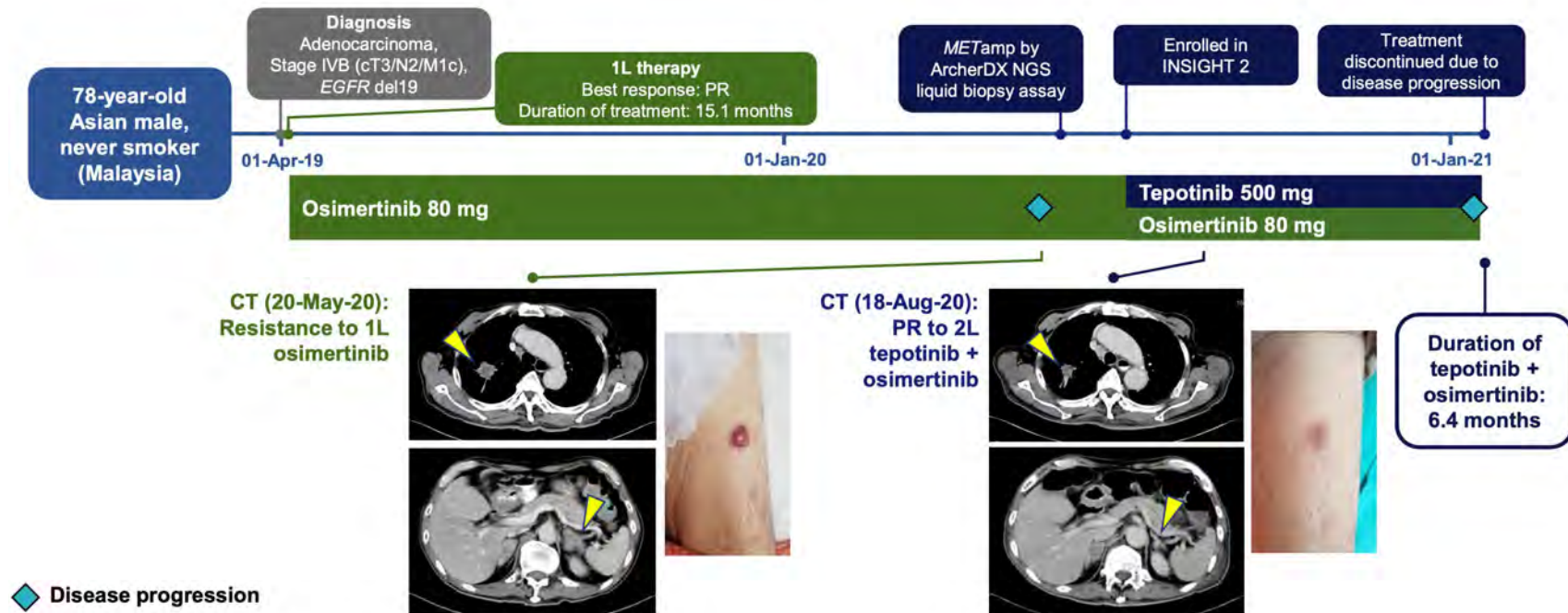
NSCLC: non-small cell lung cancer; PFS: progression-free survival.

Case courtesy of Dr Rolfo, University of Maryland School of Medicine.

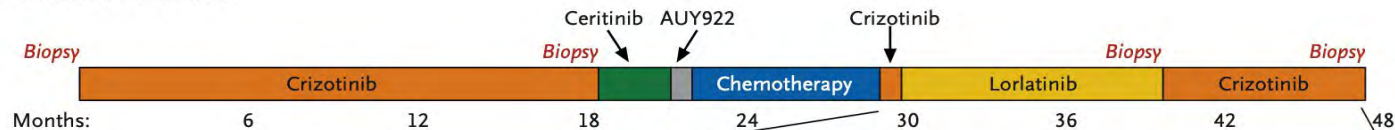
Clonal evolution of treatment resistance



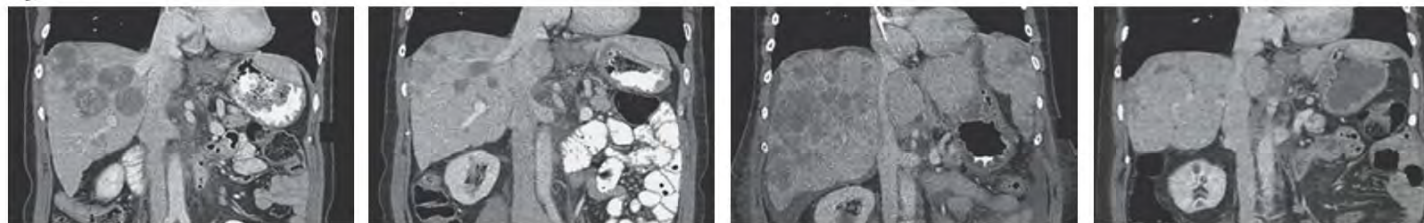
Prolonged duration of tepotinib + gefitinib after progression on erlotinib in a patient with *METamp*



A Timeline of Treatment



B Effect of Therapy

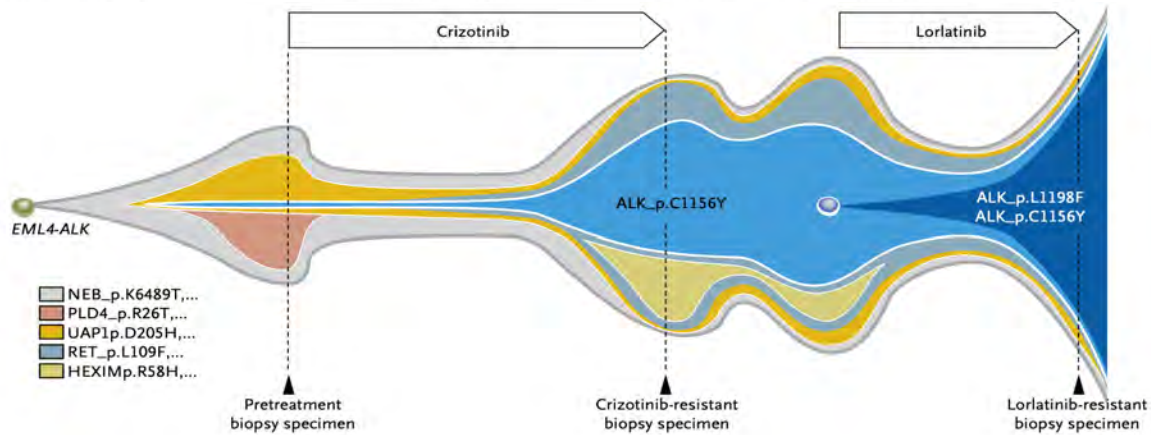


Before Lorlatinib

Response to Lorlatinib

Resistance to Lorlatinib

Response to Crizotinib



NEW KIDS ON THE BLOCK



VISION

Phase 2 trial

Single-arm study of tepotinib in stage IIIB/IV NSCLC (all histologies) with *MET* ex14 skipping mutations (Cohort A)

First-, second- and third-line therapy patients included, unless prior anti-*MET* therapy was used

Patients with active brain metastases excluded

Tepotinib in NSCLC with *MET* exon 14 skipping mutations (*MET* ex14)

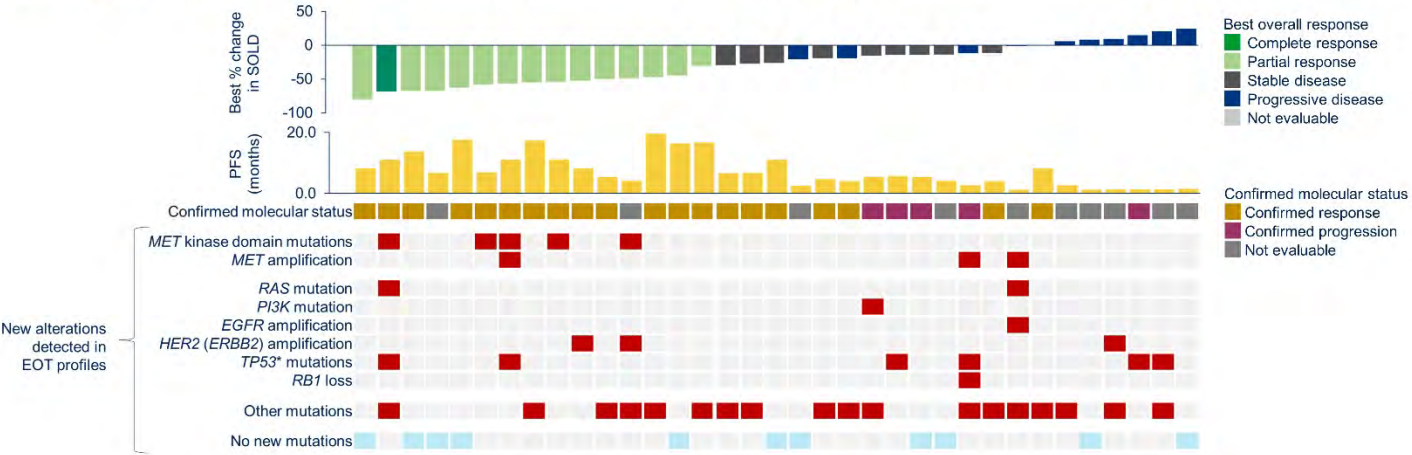
<i>MET</i> ex14 positive by:	Liquid biopsy (n=48)	Tissue biopsy (n=51)
Best overall response by RECIST 1.1 (independent review committee), n (%)		
Complete response	0 (0)	0 (0)
Partial response	24 (50.0)	23 (45.1)
Stable disease	8 (16.7)	14 (27.5)
Progressive disease	7 (14.6)	8 (15.7)
Not evaluable	9 (18.8)	6 (11.8)
ORR* n (%) [95% CI]	24 (50.0) [35.2, 64.8]	23 (45.1) [31.1, 59.7]

*ORR = Complete response + partial response

METex14 ctDNA dynamics & resistance mechanisms detected in liquid biopsy (LBx)

End-of-treatment ¹⁶

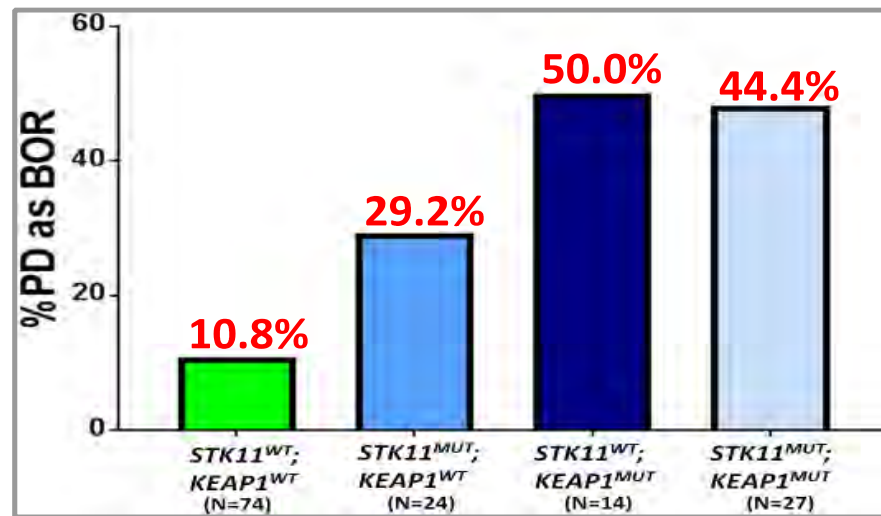
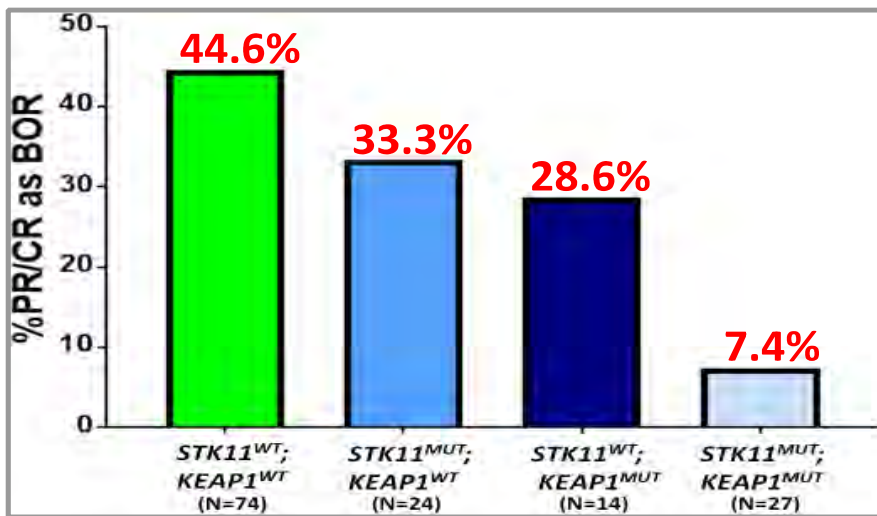
Emerging resistance mechanisms were detected in samples taken at the time of disease progression



- New alterations detected included *MET* mutation/amplification (n=7/35), *TP53* mutation/*RB1* loss (n=6/35), *EGFR/HER2* amplification (n=4/35), and *RAS/PI3K* mutations (n=3/35)
 - No high frequency bypass pathway alterations were detected
- Previously reported resistance mechanisms to *MET* inhibitors include mutations and amplifications in *RAS* and *EGFR* genes¹⁻⁴

All efficacy outcomes were investigator-assessed.
 *All non-silent *TP53* mutations included.
 1. Guo R, et al. *J Clin Oncol.* 2019;37(15):9006. 2. Awad MM, et al. *J Clin Oncol.* 2018;36(15):9069; 3. Salgia R, et al. *Can Treat Rev.* 2020;87:102022; 4. Hong L, et al. *Ther Adv Med Oncol.* 2021;13:1-16.

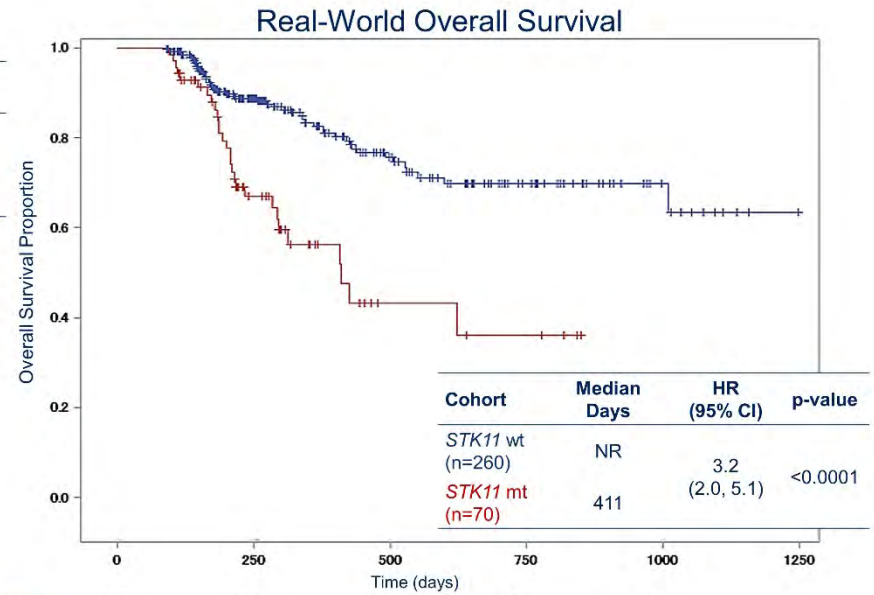
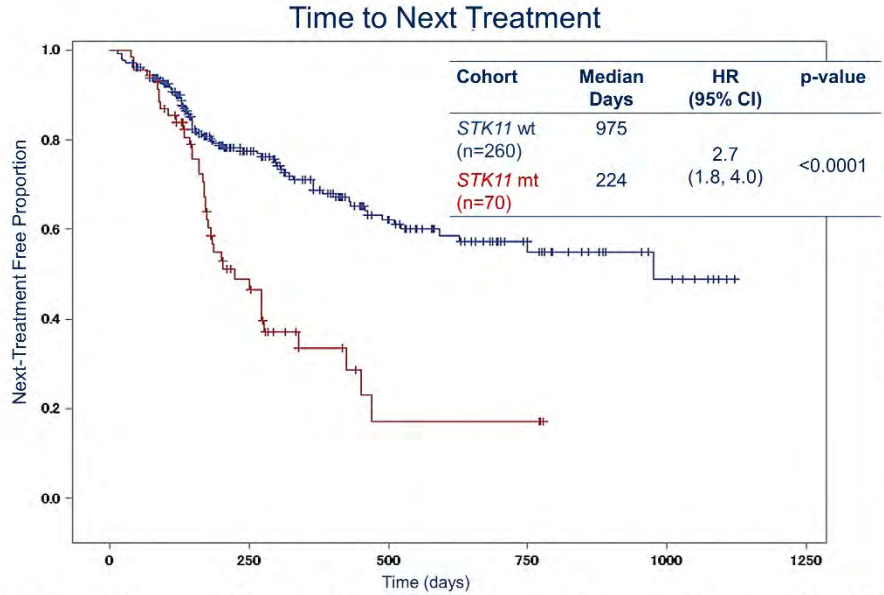
ORR with PCP in *STK11* and *KEAP1*-defined subgroups



***STK11*^{MUT} and/or *KEAP1*^{MUT} PR/CR: 21.5%, SD: 38.5%, PD 40%**

- 26/34 (76.5%) patients with primary refractory disease had *STK11* and/or *KEAP1* mutations**

Patients with *KRAS*^{G12C} and co-occurring *STK11* mutation have worse real-world outcomes with 1L ICI than those with *KRAS*^{G12C} and no *STK11* mutation



	260	146	81	41	19	8	0		260	154	71	32	11	0	
<i>STK11</i> wt	260	146	81	41	19	8	0		<i>STK11</i> wt	260	154	71	32	11	0
<i>STK11</i> mt	70	30	8	3	0				<i>STK11</i> mt	70	32	6	4	0	

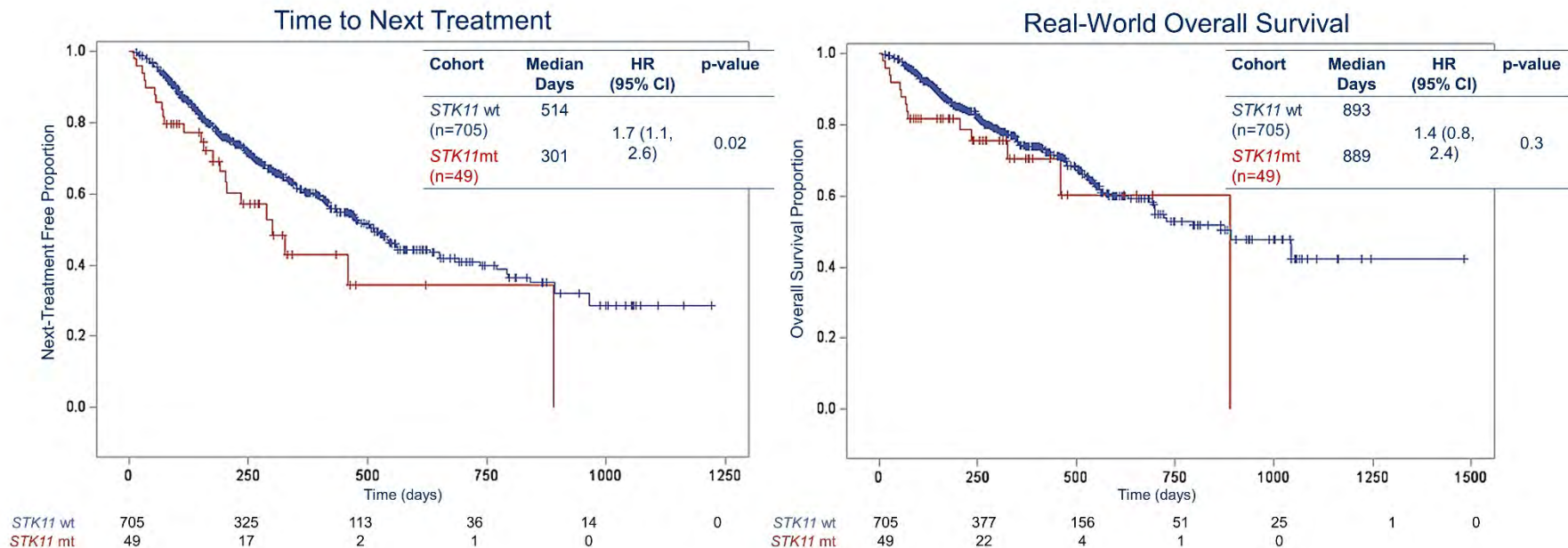
wt, wild type; mt, mutation; NR, not reached

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Shorter TTNT, but no significant difference in RW-OS with *STK11* mutation in patients with *KRAS* wild-type lung adenocarcinoma



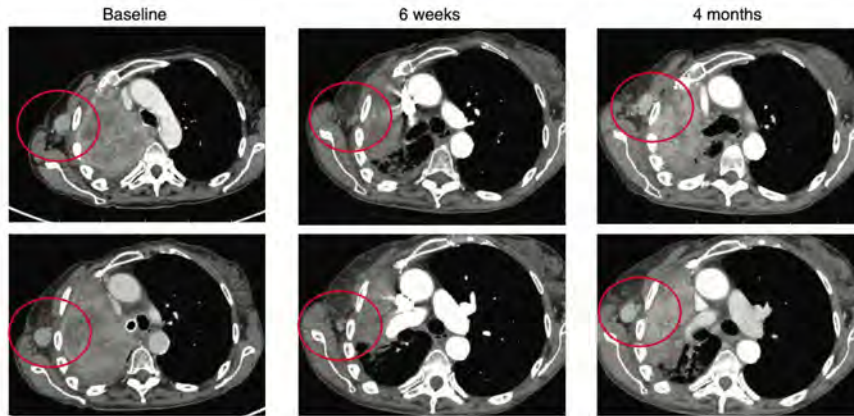
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Acquired resistance to KRASG12C inhibitor MRTX849 (adagrasib)

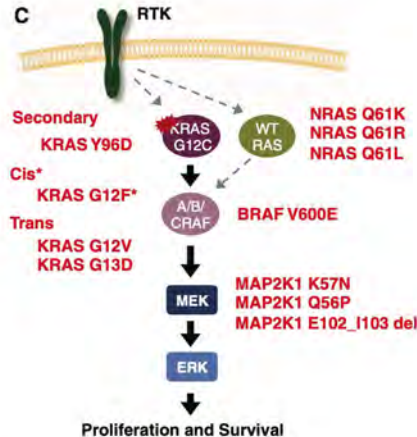


Patient with *KRASG12C* NSCLC who developed polyclonal acquired resistance to MRTX849 with the emergence of 10 heterogeneous resistance alterations in serial cell-free DNA spanning four genes (*KRAS*, *NRAS*, *BRAF*, *MAP2K1*), all of which converge to reactivate RAS–MAPK signaling.

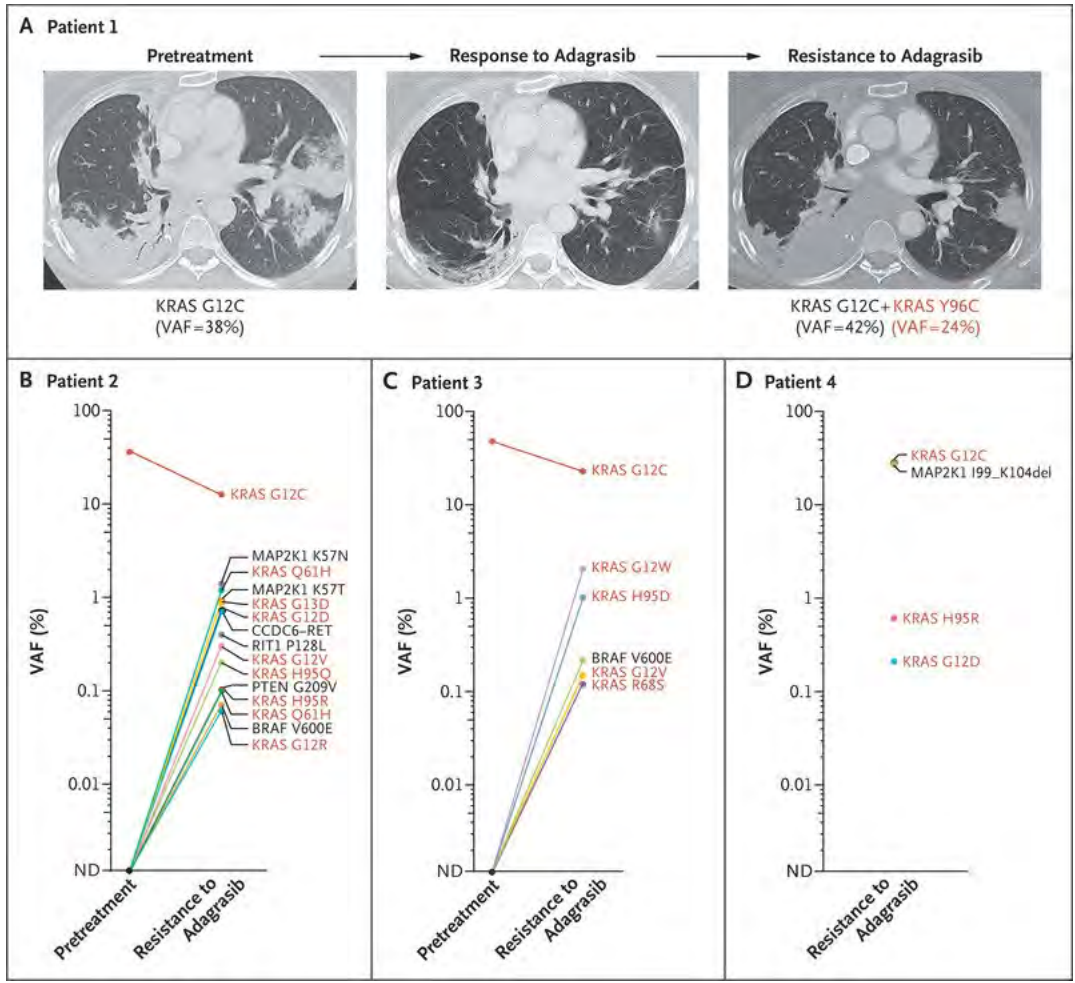
B

	Tumor		cfDNA		
	Pre-MRTX849	Pre-MRTX849	Days post-MRTX849 discontinuation:		
			0	9	51
TP53 F338fs	36.8%	0.22%	8.8%	10.1%	14.3%
KRAS G12C	21.3%	0.12%	31.7%	47.1%	24.9%
KRAS G12V	-	-	-	-	0.09%
KRAS G13D	-	-	-	0.13%†	0.04%
KRAS Y96D	-	-	0.4%	0.2%	-
NRAS Q61L	-	-	-	0.2%	-
NRAS Q61R	-	-	-	-	0.02%
NRAS Q61K	-	-	0.6%	0.6%	0.9%
BRAF V600E	-	-	0.1%	0.1%	0.5%
MAP2K1 K57N	-	-	0.05%†	-	0.3%
MAP2K1 Q56P	-	-	-	-	0.1%
MAP2K1 E102_I103del	-	-	-	0.12%†	0.2%

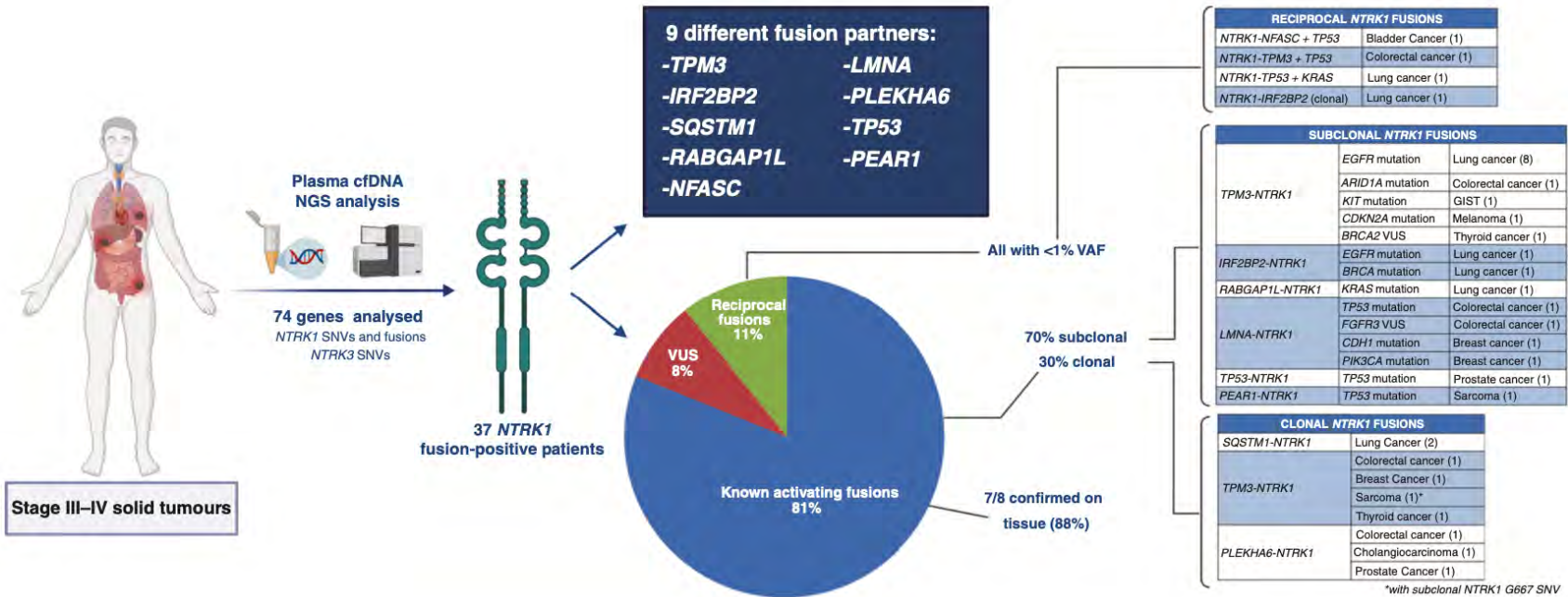
C



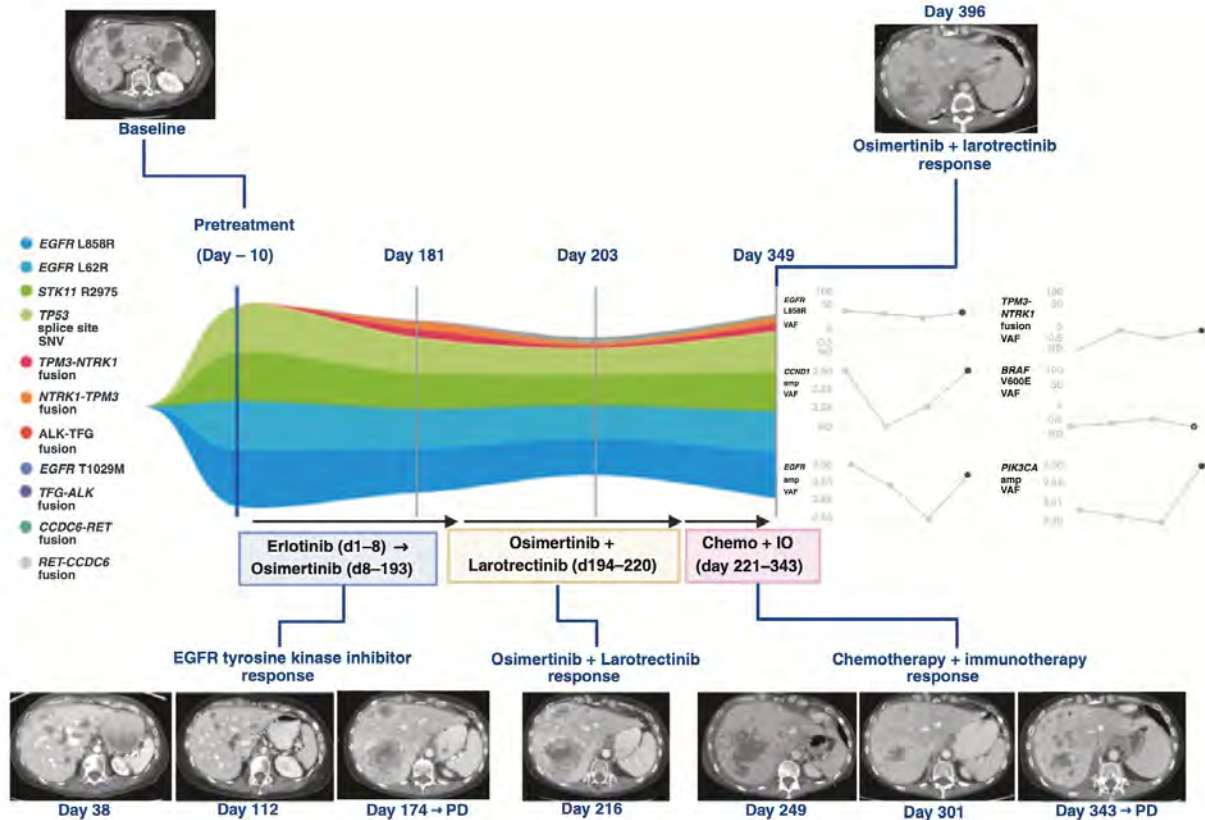
Variant allele fractions of mutations detected in the patient's serial plasma samples. †, indicates the mutations were detected by ddPCR but not by plasma next-generation sequencing

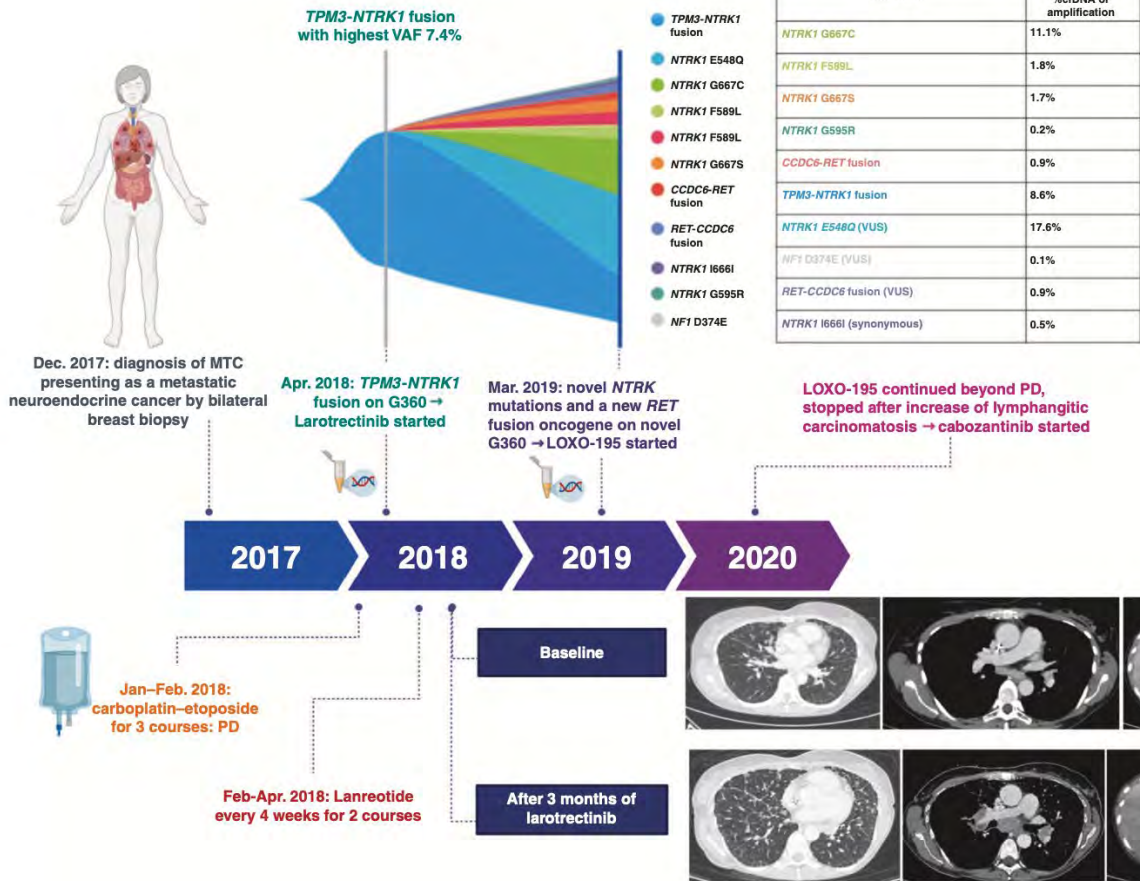


NTRK1 Fusions identified by non-invasive plasma next-generation sequencing (NGS) across 9 cancer types

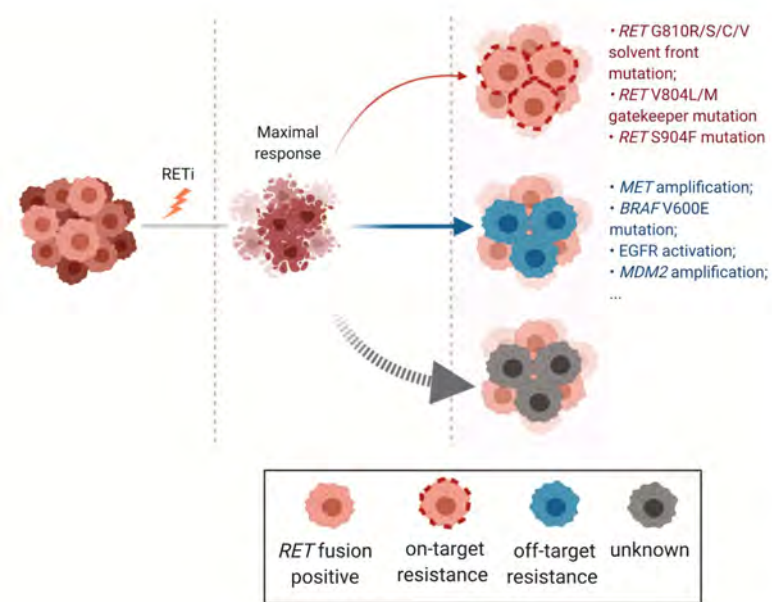


NTRK in Lung Cancer as a mechanism of resistance of EGFR



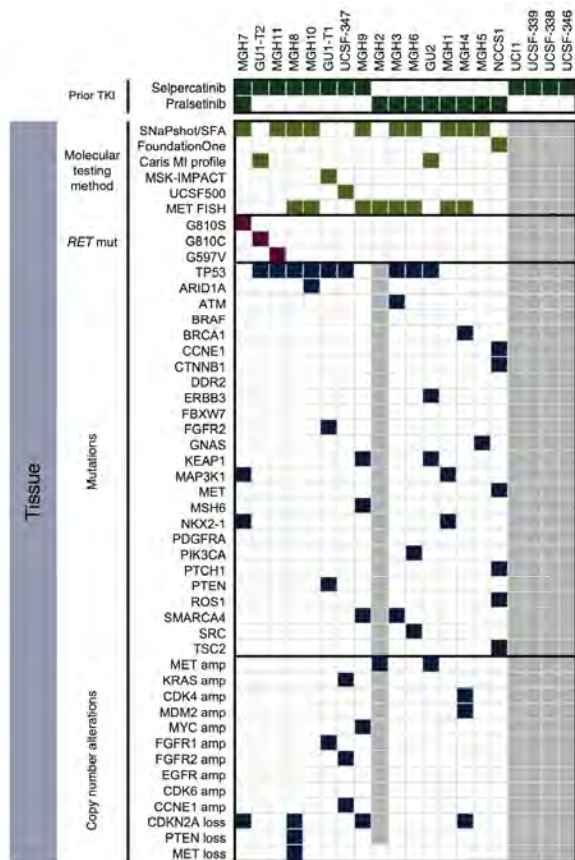


Resistance to Novel Selective RET Inhibitors in Lung Cancer.

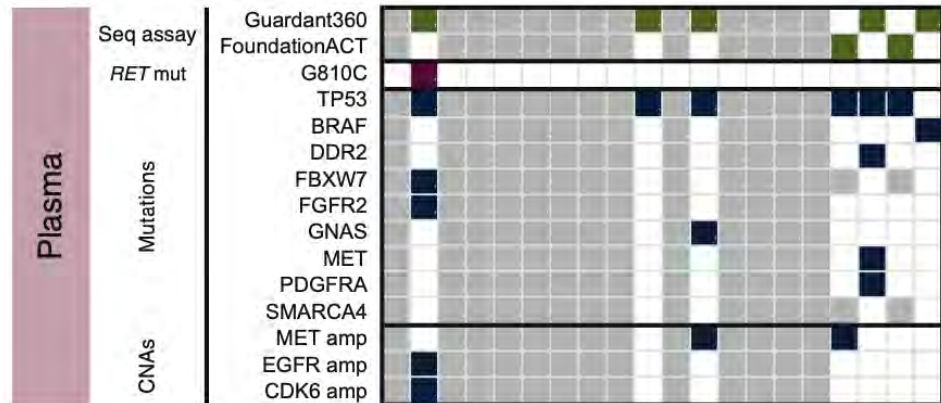


Mutation Status		Cabozantinib [112]	Vandetanib [112]	Lenvatinib [112]	Ponatinib [112]	Selpercatinib [109]	Pralsetinib [109]
Gatekeeper	V804M	4.26	5.83	5.42	0.0339	0.0559	0.0168
	V804L	3.22	6.10	10.60	0.43 [60]	0.0172	0.0018
Solvent front	G810A	0.22	2.76	0.11	0.008 [60]	-	-
	G810R	-	-	-	-	2.744	2.650
	G810S	1.05	5.47	0.67	-	0.8802	0.3906
	G810C	-	-	-	-	1.227	0.6417
Other	S904F	-	0.908 [28]	-	-	-	-
	Y806C	-	0.933 [113]	-	-	0.1744	0.2958
	Y806N	4.76	5.86	1.93	-	0.1498	0.2925
	V738A	1.20	1.05	2.35	-	0.2388	0.1775

RET fusions

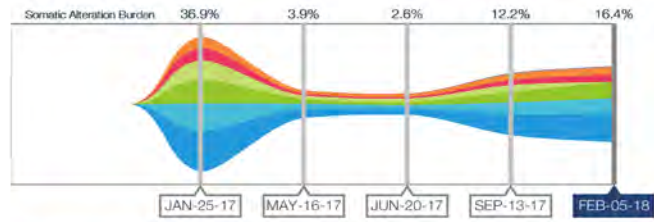


- RET inhibitor prior to biopsy
- Molecular testing method
- RET mutation
- Non-RET gene alteration
- Alteration not detected
- Testing not performed

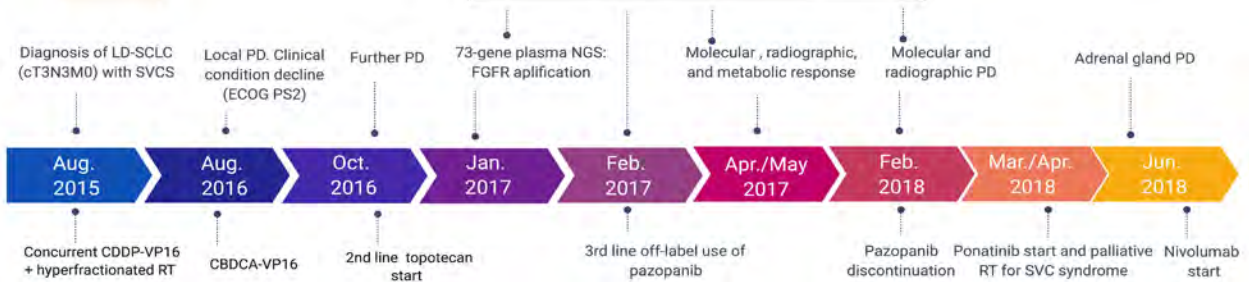


On 18 pts Acquired RET mutations were identified in two cases (10%), both affecting the RET G810 residue in the kinase solvent front. Three resistant cases (15%) harbored acquired MET amplification without concurrent RET resistance mutations, and one specimen had acquired KRAS amplification.

Thinking outside the box



Genetic alterations	% cDNA or amplification				
TP53 C176F	36.9%	3.9%	2.6%	12.2%	16.4%
RB1 R753*	22.7%	2.2%	1.3%	6.0%	7.7%
CDKN2A A57S	19.0%	2.0%	1.2%	5.3%	8.2%
PTEN S287*	8.6%	0.6%	0.4%	2.0%	2.9%
ME7 D89F*	14.7%	1.1%	1.1%	2.9%	ND
ME1 Q11E	7.1%	1.1%	0.9%	2.7%	3.2%
AR S29R	-	-	0.1%	-	-
NOTCH1	-	-	-	0.2%	ND
FGFR1 AMP	+++	ND	ND	++	++
MYC AMP	++	ND	ND	ND	ND
RAF1 AMP	++	ND	ND	ND	+
BRAF AMP	+	ND	ND	ND	ND
CDK6 AMP	ND	ND	ND	ND	+



Liquid Biopsy in Personalize Treatment

- cfDNA is a good tool fo all the patient's journey
- Make sure you use a validated platform, comprehensive
- Report every acencdotical case!
- Test your patients! At least to know the ones will not response to
Immunotherapy



Don't rush
Marty! Still
there are people
who is not
testing!

So Doc ?? Is the
future of liquid
biopsy really
cool?

Exosomes, PLTs,
CTS, multomics,
all over the place!!

I can't wait to
be there!!



Thanks



@isliquidbiopsy



Thanks



Christian.Rolfo@mssm.edu



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